COMPARATIVE STUDY BETWEEN CHEMICAL AND NON-CHEMICAL CONTROL AGAINST Sclerotium cepivorum, THE CASUAL WHITE ROT OF ONION UNDER EGYPTIAN CONDITIONS.

El-Sheshtawi, M.*; T. E. El-Gazzar** and Amany S. M. Saad*

* Plant Pathology Dept. Faculty of Agriculture, Mansoura University, Egypt.
** Vegetable and Floriculture dep. Faculty of Agriculture, Mansoura University, Egypt.

ABSTRACT

Sclerotium cepivorum one of the most destructive soilborne pathogens affecting onion and other Allium species and causing considerable damage to the host under congenial environments. In this work, it was isolated from North Egypt; many attempts were done to control onion white rot disease caused by the pathogen. Three fungicides; procimidone (sumisclex® 25), vinclozolin (Ronilan) and tolclofos-methyl (Rizolex) were used against S. cepivorum and compared with some biological non-chemical treatments. These treatments include a five antagonistic fungi, two antagonistic bacteria, three essential oils and three plant extracts. Among all fungicides tested, it was found that procymidone application was the best chemical treatment giving 100% inhibition in mycelial growth and reduction of sclerotia germination at all concentrates tested, in-vitro. While, T. harzianum was the best antagonistic fungi against the pathogen giving 82.8% reduction. On the other hand, Pseudomonas fluorescens and Bacillus subtilis have achieved complete inhibition of mycelial growth giving 100 and 94.00%, respectively and suppressed completely sclerotial germination when exposed to bacterial culture in half-strength Nutrient Broth (NB). Among all tested essential oils, it was noticed that, cinnamon (Cinnamomum zeylanicum) was the best essential oil for reducing mycelial growth giving 100% at 0.50 and 0.75%. While the cinnamon extract was the best against S. cepivorum with 94.40% inhibition in mycelial growth at 50% concentrate and completely suppressed sclerotial germination. Conversely, onion white peels extract gave the lowest rate of mycelium growth reduction giving (11.11%) and stimulated sclerotial germination than other extracts.

Keywords: chemical control, non-chemical control, S. cepivorum, onion

INTRODUCTION

Onion (Allium Cepa L.) is the most widely cultivated Allium species in Egypt. Onion production has been significantly reduced mainly in Upper Egypt due to white rot caused by Sclerotium cepivorum Berk. It has become a recurrent problem in major onion production areas all over the world (Mengistu and Seid, 1993; Mengistu, 1994). The disease is prevalent in many Allium growing regions worldwide and causes serious economic losses in onion and garlic crops (Perez et al., 1994; Crowe, et al., 1980; Andrea et al., 1996 and Pinto et al., 1998). S. cepivorum has a global importance causing serious white rot disease on Allium species like onion, garlic, shallot and leek (Coley-Smith et al., 1987 and Entwistle, 1986, 1990a). The pathogen
produces numerous small size sclerotia which aid in survival, and consider as a primary source of inoculum. On germination, the sclerotia produce mycelium which penetrates the root epidermis and invades the cortical parenchyma both intra and intercellulary causing extensive tissue degradation (Abd-El-Razik et al., 1973). Infected plants suffer water stress and usually die prematurely (Entwistle, 1990b). Management of soilborne diseases especially those that produce sclerotia is very difficult and need an integrated strategy. Crop rotation with non host crops (Banks and Edgington, 1989), soil solarization (Porter and Merriman, 1983 and Melero-Vara et al., 2000), biological control agents (Harrison and Stewart, 1988; Kay and Stewart, 1994b and Gerlagh et al., 1996), sclerotia germination stimulants (Coley-Smith and Parfitt, 1986), and composted onion waste (Coventry et al., 2002) have been tried with varying levels of success. However, no single method gave the desired level of white rot control. Fungicides are among the most effective options for white rot management. Avila De Moreno, 1991 found that vinclozolin and carbendazim applied 45 and 75 days after sowing gave the best control of the disease. Previously it has been also reported that vinclozolin and iprodione (Utkhede and Rahe, 1979), procymidone (Stewart and Fullerton, 1991; Fullerton and Stewart, 1991) gave reduction of disease incidence up to 75–95% applied as seed and soil treatment. (Melero-Vara et al., 2000) found that tebuconazole was effective in reducing the incidence and progress of the disease and in increasing the yield when applied as a clove treatment. According to(Duff et al., 2001) procymidone and tebuconazole applied as seed treatment resulted in better yields. Application of fungicides could also be integrated with other disease management components for effective control of white rot on garlic. In view of the extensive recurrent incidence of white rot in different areas of Egypt, the present studies were conducted with the objectives to (I) determine the effects of fungicides on epidemics of white rot of onion (II) determine the effects of antagonistic fungi (III) determine the effects of antagonistic bacteria (IV) determine the effects of some essential oils (V) determine the effects of some plant extracts. The efficiency of these biocontrol agents and other non-chemical control means were tested for different points of view i.e. on mycelial growth and germination of sclerotia.

**MATERIALS AND METHODS**

**Isolation, purification and identification of the causal of onion white rot disease (S. cepivorum)**

Samples of infected onion bulbs showed the typical symptoms of white rot were collected from different locations of Dakahlia and Gharbia governorates. Two methods were used to isolate the pathogen from the collected samples, i.e., by means of mycelium or sclerotia. Isolation from infected bulbs was conducted by picking off mycelial growth from diseased onion bulbs and roots according to Clarkson, et al. (2002). Where isolation from sclerotia was according to Harper and Stewart (2000) and Clarkson, et al. (2002)
Effect of certain fungal antagonists on growth of \textit{S. cepivorum} in-vitro:

Five antagonistic fungi, (\textit{T. viride}, \textit{T. harzianum}, \textit{Coniothyrium minitans}, \textit{G. virens} and \textit{Penicillium janthinallum}) were isolated from rhizosphere of healthy onion plants grown in Dakahlia governorate. Roots of plants were washed carefully with tap water to remove the adhering soil particles. The washed roots were cut into small pieces and divided into two groups. The first group was surface sterilized by immersing the root pieces in 1% Na-hypochlorite solution for 5 minutes and then washed several times in sterilized distilled water to remove any residual effect of Na-hypochlorite, while the second group was left without sterilization in order to isolate the surface organisms. The washed root pieces were dried between two sterilized filter papers, then transferred to the surface of Potato dextrose agar (PDA) amended with rose Bengal (0.003 \%) and streptomycin sulfate (0.01 \%) in Petri dishes and incubated at 25±2°C for 4-7 days. The growing fungi were individually transferred to PDA medium. Purification of fungi was carried out using single spore or hyphal tip technique. The purified fungal isolates were identified by Dept. of Plant Pathology, Faculty of Agriculture, Mansoura University. \textit{C. minitans} was obtained as a kind gift from Prof. Dr. Laszlo Vajna, Dept. of Plant Pathology, Institute of Plant Protection Research, Budapest, Hungary. The effect of these antagonistic fungi on radial growth of \textit{S. cepivorum} was studied. Each of obtained antagonistic fungi was grown on PDA for 5-7 days at (25±2°C), the fungal pathogen was grown on PDA for 5-7 days at (20±2°C) under dark condition. The antagonistic was done through using one disc (5 mm. in diameter) of the antagonist facing one disc of the pathogen on the PDA surface and relatively closed to the periphery of the plates. The untreated control treatment was done on the same medium in Petri dishes by growing one disc of the pathogenic fungus in the same place but without antagonistic disc plate. Three replicates were used. All plates were incubated at (20±2°C) in dark for 7 days after inoculation; average of radial growth (mm.) was recorded and compared with the untreated control to calculate the inhibition %

Effect of antagonistic bacterial action on radial growth of pathogen:

The inhibitory effects of \textit{Bacillus subtilis} and \textit{Pseudomonas fluorescens} on radial growth of \textit{S. cepivorum} were studied. \textit{B. subtilis} and \textit{P. fluorescens} were obtained from laboratory of organic agriculture, Agriculture Research Center (ARC) in Cairo. All pure cultures of \textit{B. subtilis} and \textit{P. fluorescens} were grown on Nutrient Agar medium (NA) for 48h. (30±2°C). The fungal pathogen was grown on PDA for 5-7 days (20±2°C). The antagonistic effects of the used bacteria on the fungal pathogen was done through streaking the antagonistic bacteria facing one disc of the pathogen on the PDA surface and relatively closed to the periphery of the plates. The untreated control treatment was done on the same medium in Petri dishes by growing one disc of the pathogenic fungus in the same place of treatments without antagonistic bacteria. Incubation was done for 7 days at (20±2°C) in dark; average of the pathogenic fungus radial growth (mm.) was recorded and compared with the untreated control %.
Antifungal activity of some essential oils on growth of *S. cepivorum*:

Three commercial essential oils of cinnamon (*Cinnamomum zeylanicum*), garlic (*Allium sativum*) and onion (*Allium cepa*) were tested for their antifungal activity at three concentrations 0.25, 0.50 and 0.75% (v/v) against the fungal pathogen. The three essential oils concentrates prepared by mixing with PDA medium, after autoclaving with 0.5% of Tween-80 (v/v) to enhance oil solubility, then the essential oil concentrates were added before solidifying, and then poured in sterile Petri dishes (9cm). Three replicates for each concentrate were used. Control treatment was done by mixing PDA with tween-80 only with no essential oils added. All plates were left for 30 min to be solidified before inoculation with 5 mm discs diameter taken from 7 days old culture of pathogen. After 7 days, average of radial growth (mm.) was recorded and compared with the untreated control %.

Antifungal activity of plant extracts on growth of *S. cepivorum*:

Three extracts from onion red peels, onion white peels and cinnamon bark were tested against the fungal pathogen. Their extracts were prepared from dry part; Onion dry peels (100g) were ground into fine powder in a high-speed micro mill. The powder were soaked in distilled water at the rate of 1:4 (w/v) then, the mixture was heated at 100°C for 30min., and filtrated through cheese cloth under a strong hand pressure. The extract was centrifuged at 12000 rpm for 30 min. and sterilized by filtering through a 0.22 μm membrane filter at 25±3°C to avoid any bacterial or fungal contamination. The extract was considered as 100% concentration. Then was mixed with PDA at 48°C to obtain concentrations of 10, 25 and 50 %, with 0.5% of Tween-80 (v/v). Cinnamon soaked in distilled water at the rate of 1:2 (w/v) then was mixed with PDA at 48°C to obtain concentrations of 0, 10, 25 and 50 % with 0.5% of Tween-80 (v/v) to enhance oil solubility, The amended media were poured into 9 cm Petri dishes (12 ml per plate). Three replicates for each concentrate were used. Control treatment (0%) was done by mixing PDA with tween-80 only with no extracts added. All plates were left for 30 min to be solidified before inoculation with 5 mm disks of the pathogen, taken from 7day old culture in the centre of each plate and then incubated at 20°C. Average of radial growth was recorded after 7 days compared with the untreated control percentage when mycelial growth covered the surface of all cultures in the control treatment. Inhibition of growth was calculated in relation to the growth in the control, according to the equation proposed by Pinto, *et al.* (1998).

Effect of some fungicides on the radial growth of *S. cepivorum* causing onion white rot:

The fungicidal effects of three fungicides were determined on the radial growth of *S. cepivorum*. The tested fungicides were procimidone (Sumisclex®25), vinclozolin (Ronilan®) and tolclofos-methyl (Rizolex®). Fungicides were added to molten autoclaving PDA medium to produce the concentrations 0.5, 1, 1.5, 2, 2.5 and 3mg /100ml. Unamended PDA considered as a control, then poured in sterile Petri dishes. Three replicates for each concentrate were used, all plates were left for 3min to be solidified
RESULTS AND DISCUSSION

1- Effect of antagonistic fungal action on radial growth of pathogen.

Table (1) show that, after 5 days of incubation P. janthinallum was the best antagonistic fungus for inhibition radial growth of the pathogen giving 79.18% when compared with the untreated control. This was followed by G. virens and C. minitans giving 57.50% and 50.00% inhibition, respectively. Whereas, T. harzianum and T. viride were not effective on S. cepivorum giving 18.75% and 0% reduction when compared with untreated control.

After 10 days of incubation, it was noticed that P. janthinallum was the best antagonistic fungus reducing radial growth of S. cepivorum giving 86.09% inhibition, followed by T. harzianum, C. minitans, G. virens and T. viride giving 76.39%, 75.65%, 73.91% and 71.74%, respectively.

After 15 days of incubation, T. harzianum, T. viride, G. virens and P. janthinallum achieved a best inhibition among the tested antagonistic fungi giving 82.69%, 80.00%, 78.08% and 76.19%, respectively in radial growth of S. cepivorum, but C. minitans gave moderate inhibition rates of 58.85% when compared with untreated control. These results are in agreement with Djonovic, et al., 2006; Karthikeyan et al., 2006; Embaby, et al., 2007 and Yang, 2007.

This high antagonistic action of T. harzianum due to mycoparasitism, produce several fungitoxic cell-wall-degrading enzymes (Chet et al.,1998), and probably also peptaibol antibiotics. These lytic enzymes, which act as fungal cell-wall degrading agents such as N-acetyl-β-D-glucosedeaminidase, chitinase, β-1,3gluocose, chitobiosidase and protease (Elad et al.,1995; Elad et al.,1998; Antal et al.,2000 and Harman et al.,2004).

Moreover, Trichoderma spp. could manufacture cynamide hydratase, rhodanese and β-cyanoalnine synthases, which play an important function in reducing the growth of pathogenic fungi ( Ezzi-Mufaddal and James, 2002).

P. janthinallum gave high reduction of mycelial growth and sclerotia germination (Table 2). This result agree with (Gudrun, et al., 2005) who reported that P. janthinallum limited the growth of S. cepivorum through secreting some enzymes such as peptidas and endoglucanase.

On the other hand, C. minitans gave moderate inhibition rates giving 58.85% of mycelial growth of pathogen and inhibition sclerotal germination of 100%. This result showed that C. minitans specialized antifungal agent that targets sclerotia of Ascomycotina and Deuteromycotina and required a host to be in vegetative stage (Lebertz, 1995). Degradation of mycelial growth and sclerotia of S. cepivorum are due to C. minitans secreting β-1, 3- glucanase and chitinase (Li Ren, et al. (2007).
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Table (1): Effect of antagonistic fungal action on radial growth of *S. cepivorum*:

<table>
<thead>
<tr>
<th>Treatment</th>
<th>After 5day R.G Inh. %</th>
<th>After 10day R.G Inh. %</th>
<th>After 15day R.G Inh. %</th>
<th>Ger. S. No.S. Inh. %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>4.00a 0.00</td>
<td>7.67a 0.00</td>
<td>8.67a 0.00</td>
<td>25.00a 83.33</td>
</tr>
<tr>
<td><em>T. viride</em></td>
<td>4.00a 0.00</td>
<td>2.17bc 71.74</td>
<td>1.73c 80.00</td>
<td>0.00d 100</td>
</tr>
<tr>
<td><em>T. harzianum</em></td>
<td>3.25b 18.75</td>
<td>2.50b 76.39</td>
<td>1.50c 82.69</td>
<td>0.00c 100</td>
</tr>
<tr>
<td><em>G. virens</em></td>
<td>1.70c 57.50</td>
<td>2.06c 73.91</td>
<td>1.90c 78.08</td>
<td>0.00d 100</td>
</tr>
<tr>
<td><em>C. minitans</em></td>
<td>2.00c 50.00</td>
<td>1.87c 75.65</td>
<td>3.57b 58.85</td>
<td>0.00e 100</td>
</tr>
<tr>
<td><em>P. janthinallum</em></td>
<td>0.83d 71.78</td>
<td>1.07d 86.09</td>
<td>2.06c 76.19</td>
<td>0.00f 100</td>
</tr>
<tr>
<td>L.S.D.</td>
<td>0.7138</td>
<td>0.5641</td>
<td>0.6967</td>
<td>0</td>
</tr>
</tbody>
</table>

*R.G=Radial growth (cm.) Inh. %= inhibition %; Ger.S. = germination of sclerotia.

2-Effect of antagonistic bacterial action on radial growth of *S. cepivorum*:

Results in Table (2) indicated that *P. fluorescens* followed by *B. subtilis* achieve the highest inhibition of radial growth during the incubation period, without any significant differences between them. After 15 days, they gave maximum reduction in radial growth by 100% and 93.46%. These results have the same opinion with (Chen and Dickman, 2005; Basha, et al. 2006; Ahmadzadeh, et al. 2007 and Antelmann, et al. 2008). They reported that *P. fluorescens* and *B. subtilis* reduced the mycelial growth of *S. cepivorum*, *R. solani* and *F. oxysporum*. This high antifungal effect of *P. fluorescens* probably related to the degradation of chitin in hyphal and sclerotial cells by several hydrolyzing enzymes (Gooday, 1990), such as endochitinase (1,4-L-poly-N-acetyl-glocosaminidase), exochitinase, chitobiosidase and/or N-acetyl-glocosaminidase) or NA Gase are exudated from this antagonistic bacteria (Tronsmo and Harman, 1993). In addition these include simple metabolites such as 2,4-diacetylphloroglucinol, phenazine-1-carboxylic acid and pyrrolnitrin [3-chloro-4-(2-nitro-3-chlorophenyl)-pyrrole], as well as the complex macrocyclic lactone, 2,3-de-epoxy-2,3-didehydro-rhizoxin. Study of the biochemistry and mechanism of formation of these metabolites has proved useful in several ways. Pyrrolnitrin is active against *Rhizoctonia spp., Fusarium spp.*, and other plant pathogenic fungi, and it has been used as a lead structure in the development of a new phenylpyrrole agricultural fungicide. (Leonardo et al., 2004).

Table (2): Effect of antagonistic bacterial action on radial growth of pathogen:

<table>
<thead>
<tr>
<th>Treatment</th>
<th>After 5day Radial growth of pathogen</th>
<th>After 10day Radial growth of pathogen</th>
<th>After 15day Radial growth of pathogen</th>
<th>Ger. S. No.S. Inh. %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>R.G Inh. %</td>
<td>R.G Inh. %</td>
<td>R.G Inh. %</td>
<td>R.G Inh. %</td>
</tr>
<tr>
<td><em>B. subtilis</em></td>
<td>2.13b 46.6675</td>
<td>2.13b 72.17</td>
<td>0.57b 93.46</td>
<td>1.00b 3.33</td>
</tr>
<tr>
<td><em>P. fluorescens</em></td>
<td>1.77b 55.8325</td>
<td>1.77b 76.96</td>
<td>0.00b 100</td>
<td>0.00c 100</td>
</tr>
<tr>
<td>L.S.D.</td>
<td>0.6693</td>
<td>0.7475</td>
<td>0.6792</td>
<td>0</td>
</tr>
</tbody>
</table>

*R.G=Radial growth (cm.) Inh. %= inhibition %; Ger.S. = germination of sclerotia.
Whereas, the high antifungal activity of B. subtilis depends on the production of antibiotic compounds, including peptides (Banerjee and Hansen 1988 and Paik et al., 1998), lipopeptides (Arima et al., 1968), phenylpropanol derivatives (Pinchuk et al., 2002), and a novel phospholipid compound (Tamechiro et al., 2002).

3-Effect of Antifungal activity of some essential oils on radial growth and germination sclerotia of S. Cepivorum.

Results in Table (3) showed that Cinnamon oil was the most effective on radial growth of S. cepivorum giving 94%, 100% and 100% reduction in mycelium growth at all concentrates 0.25%, 0.5% and 0.75%, and suppressed sclerotia formation, except 0.25% concentrate gave 100sclerotia/plat, 3.33% germinated sclerotia when exposed in 0.5%concentrate. This is in agreement with the results of many authors who reported the antifungal activity of cinnamon oil against plant pathogenic fungi (Atta-Ur-Rahman et al., 1999 and Ranasinghe et al., 2002). This antifungal activity of cinnamon oil due to presence of some active compounds such as cinnamaldehyde, eugenol, cinnamic acid and weitherhin. Also, the antifungal activity of oil can be attributed to the presence of an aromatic nucleus and phenolics of OH group, which is Known to be reactive and form hydrogen bounds with active sites target enzymes (Velluti et al., 2003)

While, onion oil and garlic oil increased the mycelial growth gradually in all concentrations, giving 9 cm growth over than the untreated control with 8%. And forming a large number of sclerotia in all concentrations giving 3.9×10^5 and 3.7×10^5sclerotia/plat.and onion oil germinated 93.33% of sclerotia followed by garlic oil giving 90% when compared with untreated control.

<table>
<thead>
<tr>
<th>Essential oils</th>
<th>Control</th>
<th>Onion oil</th>
<th>Garlic Oil</th>
<th>Cinnamon Oil</th>
<th>Ger. S.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>8.33b 0.00 830</td>
<td>9.00a -8.00 5400</td>
<td>9.00a -8.00 2753</td>
<td>0.50c 94.00 100</td>
<td>8.00 83.33</td>
</tr>
<tr>
<td>Onion oil</td>
<td>0.00c 100 0</td>
<td>8.00c 830</td>
<td>0.00c 100 0</td>
<td>0.00c 100 0</td>
<td>0.00c 100 0</td>
</tr>
<tr>
<td>Garlic Oil</td>
<td>8.33b 0.00 830</td>
<td>9.00a -8.00 5400</td>
<td>9.00a -8.00 2753</td>
<td>0.00c 100 0</td>
<td>3.33</td>
</tr>
</tbody>
</table>

Table (3): Effect of Essential oil on radial growth and on germination of sclerotia of pathogen:

4--Effect of Antifungal activity of plant extracts on radial growth and sclerotial germination of S. Cepivorum:

Data in Table (4) illustrates the highest inhibition rates of radial growth, sclerotia forming and germination which came from Cinnamon extract, which reduced the mycelial growth by 67.22%, 89.00% and 94.44% at the all tested concentrates 10%, 25% and 50%, respectively, when compared with untreated control. It suppressed forming sclerotia by 100% at
25% and 50% concentrate, and reduced germination sclerotia giving 17.77% germination. These results agree with those obtained by Wilson, et al. (1997), they reported that cinnamon extract inhibited the radial growth of Botrytis cinerea.

Onion Red peels extract gave reduction in mycelial growth at the higher concentrate of 50% giving 51.85% and formed 1 X 10^3 sclerotia /plate, but it was deformed, while, Onion White peels gave the lowest reduction of radial growth, forming sclerotia and germination at all tested concentrates respectively. These result in agreement with Coventry, et al. (2002) and Coventry, et al. (2006). They proved that application of the leaf extract of onion alone or combined with T. viride could eliminate S. cepivorum. Alternatively, Dini, et al. (2008) stated that the extracts of onion Red peel discrete antioxidant capacity which increased after boiling, although cooking methods caused significant losses of the cysteine derivatives in water.

Table (4): Effect of water plant extracts on radial growth and on germination of sclerotia of pathogen:

<table>
<thead>
<tr>
<th></th>
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<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>9.00a</td>
<td>0.00</td>
<td>72 X 10^3</td>
<td>9.00a</td>
<td>0.00</td>
<td>72 X 10^3</td>
<td>9.00a</td>
<td>0.00</td>
<td>72 X 10^3</td>
<td>25.00a</td>
<td>83.33</td>
</tr>
<tr>
<td>Onion White Scale</td>
<td>9.00a</td>
<td>0.00</td>
<td>72 X 10^3</td>
<td>9.00a</td>
<td>0.00</td>
<td>72 X 10^3</td>
<td>8.00a</td>
<td>11.11</td>
<td>70 X 10^3</td>
<td>19.33b</td>
<td>64.43</td>
</tr>
<tr>
<td>Onion Red Scale</td>
<td>9.00a</td>
<td>0.00</td>
<td>14.67 X 10^3</td>
<td>7.83b</td>
<td>12.96</td>
<td>13.33 X 10^3</td>
<td>4.33b</td>
<td>51.85</td>
<td>1 X 10^3</td>
<td>12.33c</td>
<td>41.10</td>
</tr>
<tr>
<td>Cinnamon</td>
<td>2.95b</td>
<td>67.22</td>
<td>3.33 X 10^3</td>
<td>1.00c</td>
<td>89.00</td>
<td>0.00c</td>
<td>0.50c</td>
<td>94.44</td>
<td>0.00d</td>
<td>5.33d</td>
<td>17.77</td>
</tr>
<tr>
<td>L.S.D.</td>
<td>0.1697</td>
<td>1.5473</td>
<td>0.2718</td>
<td>1.7188</td>
<td>1.0871</td>
<td>0.9414</td>
<td>2.4908</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*R.G=Radial growth (cm.)  No.S=number of sclerotia  Ger.S. = germination of sclerotia. Inh. % = inhibition %.

5-Effect of some fungicides on the radial growth and sclerotial germination of S. cepivorum causing onion White rot:

Data in Table (5) show that the highest inhibition of fungal growth came from Sumisclex®25, which reduced the mycelial growth by 100% at the maximum concentrate(3mg/100ml) and giving 93.3% to 94.4% at concentrates of 0.5,1,1.5,2 and 2.5 mg/100ml ,when compared with untreated control, while suppressed sclerotial germination at100%at all tested concentrates, respectively. Rizolex suppressed the fungal growth significantly by 93.7. % and 94.4% inhibition, at the higher concentrate (2.5, 3%) respectively, while suppressed sclerotial germination at 100% at all tested concentrates, except in case of 0.5mg/100ml concentrate. While, Ronilan did not show any effect on radial growth but suppressed sclerotial germination by 100% at all tested concentrates. These results are in agreement with those obtained by Macleod and Nielsen (1995) and Macleod and Ryan, (1997).They found that reduction in mycelial growth of S. cepivorum achieved by dicarboximides such as vinclozolin and iprodione, while Fullerton et al., (1992). Porter, et al., (1991) and Fullerton et al., (1995) found good results have been obtained with another dicarboximide, procymidone, when compared with tebuconazole.
Table (5): Effect of fungicides on radial growth and germination of sclerotia of pathogen:

<table>
<thead>
<tr>
<th>Concentration</th>
<th>Treatment</th>
<th>0.5</th>
<th>1</th>
<th>1.5</th>
<th>2</th>
<th>2.5</th>
<th>3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>9.00a 30a</td>
<td>9.00a 30a</td>
<td>9.00a 30a</td>
<td>9.00a 30a</td>
<td>9.00a 30a</td>
<td>9.00a 30a</td>
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<td>Procymidine</td>
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<td>0.60b 1b</td>
<td>0.50b 0d</td>
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<tr>
<td>Tolclofos-</td>
<td>0.97b 29b</td>
<td>0.77b 0c</td>
<td>0.57b 0c</td>
<td>0.50c 0c</td>
<td>0.57b 1.33b 0.50c 0b</td>
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<tr>
<td>methyl</td>
<td>(Rizolex)</td>
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<td>Venconazole</td>
<td>9.00a 0d</td>
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<td>9.00a 13b</td>
<td>9.00a 1.33b 9.00a 0.33b</td>
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<td>(Ronilan)</td>
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<td>0.46 0.94</td>
<td>0.29 0.11</td>
<td>0.00 0.05</td>
<td>1.54 0.33</td>
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*L.S.D.* 0.46 0.94 0.29 0.11 0.00 0.05 1.54 0.33

R.G.=Radial growth (cm.) Ger.S. = germination of sclerotia.

REFERENCES


El-Sheshtawi, M. et al.


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