RESISTANCE OF SOME EGYPTIAN BARLEY GENOTYPES TO LEAF RUST CAUSED BY *Puccinia hordei Otth.*

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ABSTRACT

One hundred and twenty barley lines as well as three commercial cultivars belonging to the Egyptian breeding program (A, B, D & E trials) were tested for resistance to barley leaf rust (Puccinia hordei Otth.) at seedling stage and at adult stage in 2005/2006 season. Under green-house conditions, a total of 55 lines showed resistant response (low infection type). These lines comprise 45.83 %, comparing with the check commercial varieties which were susceptible (high infection types). Of these lines, 20 were highly resistant. At adult stage, the evaluation was carried out at four locations differed in climatic conditions i.e. Sakha, Gemmeiza, Nubaria and Ismalyia. Rust Severity(RS), Average Coefficient of Infection (ACI) and Relative Resistance Index (RRI) were calculated. Also ,the desirable / acceptable levels of relative resistance index (RRI) were estimated. The stability parameters using the regression coefficient of the performance of each genotype under different environments (b) and the mean square deviation from linear regression (S²d) were calculated. Most of the tested lines showed susceptible responses to leaf rust. The susceptibility to leaf rust of the tested lines was the least in Ismaylia. The line No. 29 was the best one for both resistance and stability to leaf rust disease at the four locations as it had the lowest value of ACI (5.00) and the highest value of RRI (8.38) , followed by lines No. 88, 28 and 12 where the ACI ranged between (12.50 - 14.00) and the RRI ranged between (7.45 - 7.26). These materials can be used as parents in barley breeding programs for developing new disease resistant cultivars.

Keywords: Rust Severity (RS), Average Coefficient of Infection (ACI), Relative Resistance Index (RRI), Genotypes Stability.

INTRODUCTION

Leaf rust caused by Puccinia hordei Otth. is the most important disease of barley and is widely distributed wherever the crop is grown(Clifford 1985). The disease is one of the major barley diseases in Egypt as it occurs yearly causing a considerable loss in grain yield specially in the Northern areas of Delta where environmental conditions, particularly high relative humidity, is favorable for disease development. (Ghobrial et al., 1984). The use of disease - resistant barley cultivars has been an efficient method for controlling the disease and preventing yield losses. Barley yield losses may reach 30 % in susceptible cultivars due to infection by P. hordei (Griffey et al., 1994 and Whelan et al., 1997). The development of Stable barley varieties that are tolerant to different environmental stress is the ultimate goal of the national barley program . Genotype environment interaction is often described as a consistent differences among genotypes from an environment to another .Multi -location tests would test abiotic and biotic stress against existing pathogen populations. Several attempts has been made to evaluate genotype x environmental conditions (Mirza et al.,2000 and Akhtar et al.,2001).

Regression analysis is the most widely used method proposed by Finaly and Wilkinson (1963) to estimate stability and adaptability parameters for several genotypes of barley. However, the modified model of Eberhart and Russel (1966) was widely used by various investigators in many plant species. They suggested the use of an environmental index to measure environments instead of the actual yield and supposed that any deviation from the average response (regression coefficient , b = 1) can be considered a genotype environment interaction. Thus predictable response by a variety to environment would be either good or poor. In addition to regression coefficient, the mean square for deviation from regression (S 2 d) was suggested as a useful measure of specific genotype x environment interaction .

The main objective of this study was to asses the response of various barley advancing genotypes to leaf rust at seedling stage and at adult stage in four different locations in order to use those exhibiting stable resistance in the Egyptian barley breeding program.

MATERIALS AND METHODS

One hundred and twenty advanced barley genotypes were obtained from the National Breeding Program of barley, Field Crops Research Institute, (ARC), Giza, Egypt. These genotypes comprises 4 trials: A(64 lines), B(32 lines), D(16 lines) and E(8 lines). Three highly susceptible barley varieties i.e. Giza 123, Giza 126 and Giza 2000 were used as checks in each trial, (Table 1). These genotypes were evaluated for their stable resistance to leaf rust.

Green-house test

All the barley genotypes were tested in the controlled greenhouse of the Barley Diseases Research Section. Plant Pathology Research Institute , ARC, Giza in 2005/2006 season. Five seedlings of each line / cultivar were grown in 7 cm plastic pots and inoculated by a mixture of 10 mg freshly collected uredinospores and talc powder at the rate of 1:25 according to Tarvet and Cassell,(1951). After 24 hr. of incubation in dew chamber (100% relative humidity) the inoculated plants were transferred to a greenhouse benches (20 – 24 °C). Three pots were used for each line / cultivar. Plants were investigated daily for pustules eruption until pustules establishment. Infection type was recorded following the scale of 0 - 4 according to Stakman $et\ al.$, (1962) which, 0, 0; , 1 and 2 infection types are resistant while 3 and 4 infection types are considered as susceptible .

Field test

Trials were conducted at four locations representing different climatic conditions i.e. Sakha (North Delta), Gemmeiza (Middle Delta), Nubaria (West Delta) and Ismailia (East Delta). Severe natural infection with leaf rust of barley were relied upon in the mentioned locations.

At all the four locations, the barley materials were planted in two rows of 2m. long with a row distance of 25 cm between rows. A spreader rows of highly susceptible varieties were planted around the experiment and left to

natural infection with leaf rust. A randomized complete block design with three replicates was used.

Disease parameters assessment

Disease severity of leaf rust was estimated visually as a percent of leaf area covered with leaf rust pustules according to the modified Cobb's scale 0 – 100 adopted by Peterson *et al.* (1948).

Average coefficient of infection (ACI) for each entry was calculated by multiplying the following factors by disease severity percentage according to Saari and Wilcoxson (1974),

Resistant (R) = 0.2 Moderately resistant (Mr) = 0.4 Mesothetic (X) = 0.6 Moderately susceptible (Ms) = 0.8 Susceptible (S) = 1.00

To calculate the Country Average Relative Percentage Attack(CARPA) ,the highest ACI line is set as 100 and other lines are adjusted accordingly. From CARPA values , Relative Resistance Index (RRI) is calculated according the scale (0 to 9) where, "0 "denote most susceptible and "9" as highly resistant (Akhtar *et al.*, 2002):

$$RRI = \frac{(100 - CARPA)}{100} \times 9$$

The desirable and acceptable indexs for leaf rust were estimated according to Aslam (1982), where desirable index was RRI 7 and above, while acceptable index was RRI 6 or 5.

All the obtained data were subjected to analysis of variance for each of the four locations with combined analysis of variance , Snedecor and Cochran , (1967).

Stability parameters

The stability parameters namely (b) which refer to the regression coefficient of the performance of each genotype under different environments on the environmental means overall genotypes and (S $^2 \mbox{d}$) which refers to the mean square deviation from linear regression were calculated (Eberhart and Russel , 1966) .

Table (1): List of evaluated barley genotypes for resistance to leaf rust disease, program 2005/2006.

A- Trial

	A- Irial									
No.	Pedigree and Cross Name	Source ^a								
1.	G. 123									
2.	G. 126									
3.	G. 2000									
4.	Aths/Lignee686//Giza117	Scr.1 Skh 2- 2004/05								
5.	MR25-84/Att/3/Mari/Aths//Bc/7/Aramir/Arabi Abiad/6/Man/	Scr.1 Skh 3-								
	Huiz//M69/3/Apm/RI//H272/4/CP/Bra/5/Joso"S"	2004/05								
6.	Post/Copal//Gloria-BAR/Come-B/3//4/Giza117	Scr.1 Skh 9- 2004/05								
7.	Barberusse/PI382696//Gloria-BAR/Come-B/3//3/G124	Scr.1 Skh 10- 2004/05								
8.	M66-69-1/M65-94//70-221109/3/Apm/IB65/4/Glda"S"/5/CM67/	Scr.1 Skh 14-								
	Centeno//Cam/6/Api/CM67//Aths*3/7/Aths/Lignee686/4/Rhn-03/ 3/Bc/Rhn//Ky63-1294	2004/05								
9.	M66-69-1/M65-94//70-22109/3/Apm/IB65/4/Gida"S"/5/CM67/	Scr.1 Skh 17-								
	Centeno//Cam/6/Api/CM67//Athe*3/7/Lignee527/NK1272//Alanda	2004/05								
10.	M66-69-1/M65-94//70-22109/3/Apm/IB65/4/Gida"S"/5/M67/	Scr.1 Skh 18-								
	Centeno//Cam/6/Api/CM67//Athe *3/7/Lignee527/NK1272//Alanda	2004/05								
11.	M66-69-1/M65-94//70-22109/3/Apm/IB65/4/Gida"S"/5/M67/	Scr.1 Skh 19-								
	Centeno//Cam/6/Api/CM67//Athe*3/7/Lignee527/NK1272//Alanda	2004/05								
12.	M66-69-1/M65-94//70-22109/3/Apm/IB65/4/Gida"S"/5/M67/	Scr.1 Skh 20-								
12.	Centeno//Cam/6/Api/CM67//Athe*3/7/Lignee527/NK1272//Alanda	2004/05								
13.	Gloria 'S'/Copal 'S'//As46/Aths/3/Rhn-03	Scr.1 Skh 24-								
10.	Gloria 6700par 6774340/Atti3/3/Ktili 03	2004/05								
14.	BSH-/5/Alanda/4/Lignee527//Bahtim/DL71/3/Api/CM67//Mzq	Scr.1 Skh 25-								
17.	BOIT /3/Alanda/4/Eightee321//Bantim/BE/1/3/Api/OMO///M2q	2004/05								
15.	As46/Th.Unk.27//Lignee527/NK1272	Scr.1 Skh 29-								
13.	AS40/111.011k.27//Lightee327/NK1272	2004/05								
16.	As46//DeirAlla106/Strain2055/3/Cabro/Harma	Scr.1 Skh 30-								
10.	AS40//DeliAlia 100/Stralii2000/3/Gabio/Haima	2004/05								
17.	Man/4/Ball6/Pro//APDM/Dwll/ly/3/Api/CM6715/Comper229//As46/Pro/	Scr.1 Skh 37-								
17.	6/Salda	2004/05								
18.	Carbo/Gustoe	Scr.1 Skh 38-								
19.		2004/05								
19.	N-Acc4000-301-80/FB974//Allanda-01									
20	C:447/7/A ===:=/A == i: A =:= d/C/NA==/L b ::=/NACO CO/O/A ===/D4//	2004/05								
20.	Giza117/7/Aranir/Arabi Abiad/6/Man/Huiz/M69-69/3/Apm/R1//	Scr.2 Skh 3-								
0.4	H272/9/CP/Bra/5/Joso 'S'	2004/05								
21.	Avt/Attiki/3/Giza121/Pue/4/Giza117 4,5B	Scr.2 Skh 5-								
		2004/05								
22.	SLB09-85/4/Baca 'S'/3/AC253//CI08887/C105761	Scr.2 Skh 11- 2004/05								
23.	SLB09-85/4/Baca 'S'/3/AC253//CI08887/C105761	Scr.2 Skh 12- 2004/05								
24.	Arizona5908/Aths/Lignee640/6/Giza121/Cl06248/4/Apm/lB65//11012- 2/3/Api/CM67//Ds/Apro/5/Aths	Scr.2 Skh 13- 2004/05								
25.	Arizona5908/Aths/Lignee640/6/Giza121/Cl06248/4/Apm/IB65//11012-	Scr.2 Skh 14-								
_0.	2/3/Api/CM67//Ds/Apro/5/Aths	2004/05								
26.	Barberousse/Pl382696//Gloria-BAR/Come-B/3/Giza117	Scr.2 Skh 16-								
		2004/05								
27.	M126/CM67//As/Pro/3/Lignee527/Arar/4/Giza124	Scr.2 Skh 17- 2004/05								

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Cont. Table (1)

COI	nt. Table (1)	
28.	M66-69-1/M65-94//70-22109/3/Apm/IB65/4/Glda 'S'/5/CM67/	Scr.2 Skh 22-
	Centeno//Cam/6/Api/CM67Aths*3/7/Lignee527/NK1272//Aanda	2004/05
29.	M66-69-1/M65-94//70-22109/3/Apm/IB65/4/Glda 'S'/5/CM67/ Centeno//Cam/6/Api/CM67Aths*3/7/Lignee527/NK1272//Aanda	Scr.2 Skh 24- 2004/05
30.	M66-69-1/M65-94//70-22109/3/Apm/IB65/4/Glda 'S'/5/CM67/	Scr.2 Skh 27-
00.	Centeno//Cam/6/Api/CM67Aths*3/7/Lignee527/NK1272//Alanda	2004/05
31.	ACSAD68/3/Arr/Esp//Alger/Ceres362-1-1	Scr.2 Skh 36- 2004/05
32.	Alanda-01//Gerbel/Hma/3/Gloria 'S'/Colo 'S'//Teran78	Sha.BYTM-13- 2004/05
33.	Alanda-01/4/W12291/3/Api/CM67//L2966-69	Sha.BYTM-19- 2004/05
34.	U.Sask.1766/Api//Cel/3/Weeah/4/Gaiza121/Pue	Sha.BYTM-20- 2004/05
35.	Sen 'S'/Lignee527	Sha.BYTM-23- 2004/05
36.	CABUYA/PETUNIA1//CIRU	Sha.CIMMYT-3 2004/05
37.	PETUNIAI/CALI92//BLLU	Sha.CIMMYT-15 2004/05
38.	BLLU/PETUNIA1//CABUYA	Sha.CIMMYT-16 2004/05
39.	Lignee527//Bahtim/DL71/3/Api/CM67//Mzq/5/Ager//Api/CM67/3/Cel/WI 2269//Ore/4/Hamra01	Sha.BYTL-2 2004/05
40.	Moroc9-75//W12291/W12269	BON-L 37 2004/05
41.	Alanda/Zafraa//Gloria 'S'/Copal 'S'	Sha.BYTL-21 2004/05
42.	Deir Alla106//DL71/Strain205/3/zDL529/4/Arar/Lignee527	Sha.BYT.L-69 2004/05
43.	Rhn-03//Lignee527/NK1272/3/Lignee527/Chn-01//Alanda/4/Giza121/ Pue//79An/Mn	Sha.BYT.L-81 2004/05
44.	Harmal	Sha.BYT.L-90
45.	Rihane-01/3/As46/Aths*2//Aths/Lignee686	Sha.BYT.L-96
46.	As46/Aths/3/Giza121/Pue//79n/Mn/5/Khafour/4/Rhn.03/Lignee527/NK12	Early 17-
47.	7 2/Lignee527/Chn-01//Alndra AwBlack/Aths//Arar/3/9C1279-07/Roho/7/F6-4-Kf/6/Man/Huz//M69-	2004/05 Early 23-
47.	69/3/Apm/RI/H272/4/CP/Bra/5/Josos	2004/05
48.	Alndra//Lignee527/Arar/4/TunLB923137//Arar19-3/W12291	Early 26- 2004/05
49.	Giza121/C106248/4/Apm/IB65//11012-2/3/api/Cm67// DS/Apro/5/Srs- 04/6/Cen/Bglos	Early 38- 2004/05
50.	MR25-84/Att*2//Mari/Aths*2-02 (Sel. A-22)	Early 47- 2004/05
51.	National Check	Heat.t 1-2004/05
52.	U.Sask.1766/Api//Cel/3/Weeah/4/Arar/5/As46//Deir Alla106/Strain205	Heat.t 6-2004/05
53.	BKFMaguelone1604/Atem//ER/Apm/3/Lignee640/Lignee686/4/Nainaa	Heat.t 55-
54.	Rihane-03//Lignee527/Aths	2004/05 Heat.t 59-
55.	ACSAD1182/5/Arizona5908/Avt/Attiki/3/S.T.Barley/4/Aths/Lignee686	2004/05 ACSAD -10 - 2004/05
56.	Manal/3/Lignee527/NK1272//JLB70-63/4/Barjouj	Segr. 3 – 2004/05
57.	Barjouj/5/AwBlack/Aths//Arar/3/9Cr279-07/Roho/4/Aths	Segr. 7 – 2004/05

Mostafa, Nabila A. et al.

Cont. Table (1)

58.	Aths/Lignee686//Orge905/Cr289-53-2/3/UC566/Arbayan-01//M83- 194Ras*32	Segr. 2004/05	11	_
	UC566/Arbayan-01//M83-194Ras*32/5/AwBlack/Aths//Arar/3/9Cr279- 07/Roho/4/DD-14/Ran-03	Segr. 2004/05	12	
60.	Arizona5908/Aths//Avt/Attiki/3/S.T.Barley/4/Aths/Lignee686/5/ AwBlack/Aths//Arar/3/9Cr279-07/Roho/4/Aths	Segr. 2004/05	13	_
	Arizona5908/Aths//Avt/Attiki/3/S.T.Barley/4/Aths/Lignee686/5/ AwBlack/Aths//Arar/3/9Cr279-07/Roho/4/Aths	Segr. 2004/05	14	
62.	Arizona5908/Aths//Avt/Attiki/3/S.T.Barley/4/Aths/Lignee686/5/Giza126	Segr. 2004/05	19	-
63.	Arizona5908/Aths//Avt/Attiki/3/S.T.Barley/4/Aths/Lignee686/5/Katara	Segr. 2004/05	25	_
64.	Arizona5908/Aths//Avt/Attiki/3/S.T.Barley/4/Aths/Lignee686/5/CaiMr	Segr. 2004/05	27	_

Table, 1(cont.)

B- Trial, 2005/2006

No.	Pedigree and Cross Name	Source ^a
65.	G. 123	
66.	G. 126	
67.	G. 2000	
68.	Avt/Attiki//Aths/3/Giza121/Pue/4/Line366-13-2	A-11 2004/05
69.	Avt/Attiki//Aths/3/Giza121/Pue/4/Line366-13-2	A-12 2004/05
70.	Ssn/Bda//Arar/3/C.C89	A-13 2004/05
71.	Ssn/Bda//Arar/3/C.C89	A-14 2004/05
72.	CAPA-BAR/3/API/BM67-B//MZQ/4/C114032/5//6/Sawsan/Lignee640	A-16 2004/05
73.	Monroe/Esperanza//Quina/3/Orge905/Cr.289-53-2	A-16 2004/05
74.	Ssn/Sllo/3/Amapa/Cota//Glori-BAR-/Copal/4/Orge905/Cr.289-53-2	A-20 2004/05
75.	Ssn/Sllo/3/Amapa/Cota//Glori-BAR-/Copal/4/Orge905/Cr.289-53-2	A-21 2004/05
76.	Aths/Lignee686/ACSAD618	A-27 2004/05
77.	Aths/Lignee686/5/Apm/RL/4/Api/EB489-8-2-15- 4//por/U.Sask1766/3/Cel/C1	A-28 2004/05
78.	Aths/Lignee686///Asse/Jaidr	A-30 2004/05
79.	Alnda//Lignee527/Arar/4/Coho/zy//Masurka/3/Alanda/5/ TunLB-932137/Noor17	A-31 2004/05
30.	Enir/Nacta//Ast907/3/Avt (9-9)	A-32 2004/05
31.	kenyaResearch/Belle//As46/Aths*2/3/Arar/19-3//W12291	A-33 2004/05
	Enir/Nacta//Ast907/3/Avt (9-9)	A-35 2004/05
33.	Rhn-03//Lignee527/As45	A-38 2004/05
34.	National Check	A-42 2004/05
35.	80-5145/Hma-01/3/Arar/19-83/W12291	A-43 2004/05
36.	Agir8//Alnada/Zafraa	A-44 2004/05
37.	Arar/Lignee527//Arar/Rhn-03	A-45 2004/05
38.	Alnada//Lignee527/Arar	A-47 2004/05
39.	Alnada-01/4/W12291/3/Api/CM67//L2966-69/5/Rhn-08/3/	A-49 2004/05
	DeirAlla106//DL71/Strain205	
90.	Arar//Hr/Nopal/3/Alnada-01/Alnada01	A-51 2004/05
91.	Cen/Bglo '5'/Baca 'S'/3/AC253//C108887/C105761/4/Mari/Aths*2//M-Att-	A-55 2004/05
	73-337-1	
92.	CopmCr229//As46/Pro/3/Srs/4/RWA-M47	A-56 2004/05
93.	Alnada/Hamra//Alnada-01	A-59 2004/05
94.	Arbayan/NK1272/4/Arar/3/Mari/Aths*2//M-Att-73-337-1	A-61 2004/05
95.	QB813-2/4/Hma-02//11012-2/CM67/3/Arar	A-62 2004/05
96.	QB813-2/3/Alnada-01//Ssn/Lignee640	A-63 2004/05

Table 1 (cont)

D- Trial, 2005/2006

No.	Pedigree and Cross Name	Source ^a
97.	G. 123	
98.	G. 126	
99.	G. 2000	
100.	Alnada/Hamra//Alnada01	B-5 2004/05
101.	Rihane/Giza123 (1925)	B-8 2004/05
102.	Rihane/Giza123 (1925)	B-9 2004/05
103.	Aths/Lignee86//ACSAD68	B-10 2004/05
104.	AthLignee86//ACSAD410	B-12 2004/05
105.	Nigrate/5/W12198/4/Attiki//Avt/Toi/82/Vt (Sel.2.2)	B-15 2004/05
106.	80-5145/Hma-01/3/Arar/19-3//W12291	B-19 2004/05
107.	Malouk//Aths/Lignee686	B-20 2004/05
108.	Alanda/3/C108887/C105761//Lignee640/4/Alnada/Lossalka	B-22 2004/05
109.	Alanda-02/4/Arizona5908/Aths//Asse/3/F208-	B-23 2004/05
	74/5/Alanda/3/C108887/C105761//Lignee640	
110.	Lignee527/NK1272//Alanda	B-24 2004/05
111.	CL10114/Attiki//NK1272/3/Mzq/C103909-2//Aths	B-26 2004/05
112.	Giza124/7/Man/Huiz//M-69/3/Apm/RI//H272/4/CP/Bra/5/Joso S'/6/Chn-01/W12291	B-27 2004/05

Table 1 (cont)

E- Trial, 2005/2006

No.	Pedigree and Cross Name	Source ^a
113.	G. 123	
114.	G. 126	
115.	G. 2000	
116.	Aths/Rihane-01//Sawsan/Lignee640	D-3 2004/05
117.	Sawsan/Badia//Arar/3/M84-76 Bon//Jo/Yrk/3/Galt//As46/4/	D-6 2004/05
	Hj34*80/Astrix/5/Aths	
118.	Giza117/3/W12197/CI 13450//Arar	D-7 2004/05
119.	Arizona5908/Aths//Lignee640/4/WI 2291/3/Api/CM67//L2966-69	D-10 2004/05
	/6/M64-76/Bon//Jo/York/3/MS/Galt//AS46/4/Hij34-80/Astrix/5/Aths	
120.	Arrivat/3/Arizona 5908/Aths//Lignee640	D-11 2004/05

^aBarley Research Section . FCRI, ARC , Giza , Egypt

RESULTS AND DISCUSSION

A total of 120 advanced barley genotypes as well as three commercial cultivars i.e. Giza 123, Giza 126 & Giza 2000 were tested against leaf rust pathogen (*Puccinia hordei*) at seedling stage under greenhouse conditions, as well as under four locations representing different climatic conditions.

Seedling test

This evaluation was conducted using artificial inoculation with uredinospores mixture of the pathogen races identified in 2005/ 2006 (Table 3). Out of 120 lines/ cultivar , 55 were resistant showing infection types ranged between 0 to type 2 comparing with the three commercial checks which showed susceptible infection types (3 and 4). These lines comprised 45.83% of the total barley lines. The lines No. 4, 10, 15,28, 43, 48, 58, 61, 62, 64, 78, 86, 87, 90, 91,94, 101, 103, 104 and 109 were highly resistant lines because they showed zero infection types. Seedling tests against barley leaf

rust isolates can give preliminary information about the level of resistance in the breeding germplasms. Similar results were obtained on other barley genotypes by El Sayed *et al.* (1991) Nooman *et al.* (1992) and Czembor and Bladenopoulos (2007).

Field test

Analysis of variance.

To assess the resistance of the tested genotypes, the combination of the four locations was used to carry out analysis of the studied barley genotypes. Data in Table (2) reveal analysis of variance of the tested genotypes across different environments. The four locations differed considerably concerning average coefficient of infection (ACI). Analysis of variance (Table,2) showed that there were highly significant differences between environments and significant genotype x location interactions. Highly significant differences among genotypes were detected, indicating the presence of genetic variability among these genotypes. A number of genotypes showed a genotype x location interactions for qualitative resistance, indicating that these entries may carry race-specific resistance genes. Park,(2003) reported that Puccina hordei is characterized by large genetic variability and the pathogen is able to overcome any R-gene rapidly. Based on this fact, the best strategy for barley breeders to control this pathogen is to increase the level of partial resistance or different other sources of resistance (Niks et al., 2000). El- Marakby et al. (1986) found that all the studied characters of cotton genotypes showed highly significant mean squares for environments, varieties and genotype environment interaction.

Table (2): Combined analysis of variance over locations for average coefficient of infection (ACI) of barley genotypes to leaf rust.

i doti			
Sources of variation	Degrees of freedom	Mean Square	F value ^a
Rep.(Location)	8	18.8438	
Location (L)	3	117972.500	20947.57**
Variety (V)	119	1525.4940	270.87**
L×V	357	633.3670	112.46**
Error	952	5.6318	

^aF value is significant at P < 0.01

Regarding to the data in Table (3) ,the average coefficient of infection revealed that the tested genotypes showed different levels of (ACI) ranged between 5.00 to 72.50. On contrast of seedling reactions, most of the evaluated genotypes showed susceptibility to leaf rust under all locations which exhibited the greatest values of ACI. However, some of them showed reactions ranged between moderate resistance in some locations especially in Ismalyia location and susceptible reactions in others. Leaf rust severity was lower at Ismalyia than the other Locations. Also, relative resistance index (RRI) has been considered a good criterion, since the highest values of RRI were associated with disease resistance. The genotype No. 29 exhibited the lowest value of ACI (5.00) consequently the highest value of relative

resistance index (8.38). However, the genotype No. 88 occupied the second rank which showed ACI value 12.50 and RRI value 7.45, followed by lines No,s 28 and 12 (ACI, 13.50 & 14.00 and RRI,7.32 & 7.26 respectively). The genotypes No,s 9, 13 and 101 showed susceptibility up-to 17 (ACI). While the others showed high ACI. Martinez et al. (2001) reported that disease severity describes the amount of rust disease and the amount of damage on the infected plants, and that it can be used as a proper parameter for evaluating the resistance of genotypes. Prescott and Saari (1975) reported that genotypes with average coefficient of infection less than 5 suggested the presence of adequate resistance, while genotypes having values between 5 and 10 suggested reasonable level of resistance. Also, genotypes having values greater than 10 suggested that genotypes in this class have less adequate resistance and should be improved or discarded. As the obtained results indicated the lack of resistant genotypes, so it is recommended to search for new sources of resistance through another host-pathogen interactions tests. These findings are in agree with Nabila, Mostafa, and Ahmed (2005).

Table (3): Reaction of barley genotypes to leaf rust in seedling stage (artificial infection) and adult stage (natural infection) during 2005 / 2006 growing season.

	Adult plant reaction**									
No.	Seedling	Location	ı / Leaf	rust seve		(1)	(2)	(3)RRI	(4)	(5)
NO.	reaction*	Gemmeiza	Sakha	Nubaria	Ismaylia		CARPA	(3)KKI	В	S ² d
1	4	70 S	60 S	60 S	15 S	51.25	70.68	2.63	1.078	157966
2	3	80 S	80 S	70 S	40 S	67.50	63.79	3.26	0.908	27.715
3	4	50 S	50 S	70 S	15 S	46.25	63.79	3.26	0.640	435.860
4	0	60 S	80 S	60 S	20 S	55.00	75.86	2.17	1.239	9.651
5	2	30 Ms	60 S	40 S	10 Mr	32.00	44.14	5.03	1.048	190.100
6	4	40 S	50 S	40 S	10 Mr	33.50	46.21	4.84	1.003	16.521
7	4	40 S	50 S	30 S	20 S	35.00	48.27	4.65	0.612	16.497
8	3	40 S	70 S	50 S	5 Mr	40.50	55.86	3.97	1.379	69.413
9	1	20 Ms	30 S	30 S	10 Mr	20.00	27.58	6.51	0.514	78.527
10	0	10 Mr	50 S	30 S	30 S	28.50	39.31	5.46	0.432	305.326
11	2	50 S	50 S	30 S	20 Mr	34.50	47.58	4.71	0.823	46.282
12	1	15 Mr	20Ms	30 S	10 Mr	14.00	19.31	7.26	0.195	156.233
13	3	30 Mr	30Ms	40 S	10 Mr	20.00	27.58	6.51	0.341	251.364
14	4	15 Mr	50 S	40 S	10 Mr	25.00	34.48	5.89	0.798	280.621
15	0	15 Mr	60 S	30 S	10 Mr	25.00	34.48	5.89	0.952	410.589
16	4	30 Mr	40 S	40 S	20 Ms	27.00	37.24	5.64	0.494	107.306
17	3	30 S	50 S	50 S	15 Mr	34.00	46.89	4.78	0.915	140.689
18	1	80 S	70 S	50 S	30 S	57.50	79.31	1.86	0.958	144.197
19	1	70 S	60 S	70 S	20 Ms	54.00	74.48	2.29	1.137	253.169
20	1	20 Ms	50 S	20 S	5 Mr	22.00	30.34	6.27	0.873	143.003
21	4	30 S	40 S	30 S	5 Mr	25.50	35.17	5.83	0.787	8.623
22	3	30 S	60 S	40 S	10 Mr	33.50	46.20	4.84	1.046	189.335
23	4	30 S	60 S	40 S	10 Mr	33.50	46.20	4.84	1.048	190.100
24	3	40 S	60 S	50 S	5 Mr	38.00	52.41	4.28	1.217	66.753
25	2	40 S	60 S	60 S	5 Mr	40.50	55.86	3.97	1.226	184.903
26	1	30 Ms	40 S	40 S	10 Mr	27.00	37.24	5.64	0.724	121.750
27	1	40 S	40 S	30 S	10 Ms	29.50	40.68	5.34	0.908	27.716
28	0	15 Mr	20Ms	30 S	5 Mr	13.50	18.62	7.32	0.238	166.761

Cont. Table (3)

Cont. Table (3)										
	Adult plant reaction Location / Leaf rust severity %								T	1
No.	Seedling response	Location Loc		rust seve	lsmaylia	ACI	CARPA	RRI	b	S²d
29	3	15 Mr	10 Mr	10 Ms	5 Mr	5.00	6.89	8.38	0.604	20.976
30	3	40s	70 S	40 S	10Mr	38.50	53.10	4.22	1.327	36.416
31	3	50 S	70 S	30 S	5 Mr	38.00	52.41	4.28	1.420	46.530
32	4	50 S	70 S	30 S	10 Mr	38.50	53.10	4.22	1.377	51.471
33	3	80 S	50 S	30 S	15 Mr	41.50	57.24	3.85	1.191	507.312
34	3	80 S	60 S	40 S	5 Mr	45.50	62.76	3.35	1.442	341.261
35	3	80 S	70 S	40 S	5 Mr	48.00	66.20	3.04	1.604	247.334
36	-	20 Ms	80 S	40 S	20 S	39.00	53.79	4.16	0.951	646.416
37	4	30 S	80 S	50 S	20 S	40.50	55.86	3.97	1.057	298.615
38	2	40 S	70 S	60 S	10 Mr	43.00	59.31	3.66	1.345	161.638
39	2	70 S	60 S	60 S	10 Mr	48.00	66.20	3.04	1.358	212.095
40	3	90 S	60 S	40 S	20 S	52.50	72.41	2.48	1.133	784.233
41	1	10 Mr	50 S	40 S	5 Mr	24.00	33.10	6.02	0.832	413.926
42	1	40 S	70 S	40 S	10 Mr	38.50	53.10	4.22	1.327	36.46
43	0	50 S	50 S	50 S	50 Mr	42.50	58.62	3.72	1.113	104.687
44	2	30 Ms	60 S	20 S	10 Mr	27.00	37.24	5.64	1.030	181.243
45	3	30s	40 S	70 S	10 Mr	36.00	49.82	4.51	0.808	597.892
46	3	60 S	60 S	30 S	10 Mr	38.50	53.28	4.20	1.273	105.699
47	1	30 Ms	60 S	20 S	40 S	37.00	51.21	4.39	0.190	425.919
48	0	80 S	60 S	60 S	20 S	55.00	76.12	2.14	1.034	241.935
49	3	40 S	40 S	40 S	10 Mr	31.00	42.90	5.13	0.840	58.911
50	2	30 S	60 S	60 S	5 Mr	38.00	52.59	4.26	1.167	286.912
51	1	40 S	60 S	40 S	10 Mr	36.00	49.82	4.51	1.088	9.360
52	3	80 S	70 S	30 S	30 S	52.50	72.66	2.46	0.940	403.594
53	3	60 S	60 S	30 S	30 S	55.00	76.12	2.15	0.661	147.438
54	4	80 S	60 S	60 S	20 S	55.00	76.12	2.15	1.034	241.935
55	1	50 S	80 S	40 S	5 Mr	43.00	59.51	3.64	1.591	30.569
56	3	40 S	50 S	40 S	30 S	40.00	55.36	4.02	0.393	4.227
57	1	30 Ms	70 S	40 S	20 S	38.50	53.29	4.20	0.828	339.151
58	0	40 S	70 S	40 S	5 Mr	38.00	52.59	4.26	1.370	33.588
59	2	50 S	60 S	30 S	30 S	42.50	58.82	3.70	0.603	92.620
60	4	60 S	70 S	50 S	10 Mr	48.50	67.13	2.96	1.453	16.681
61	0	80 S	80 S	50 S	30 S	60.00	83.04	1.52	1.120	110.109
62	0	60 S	70 S	50 S	10 Ms	47.00	65.05	3.14	1.376	11.644
63	1	60 S	70 S	40 S	20 S	47.50	65.74	3.08	1.062	41.121
64	0	80 S	70 S	50 S	10 Ms	54.50	75.43	2.21	1.493	184.406
65	4	60 S	60 S	50 S	30 S	50.00	69.20	2.77	0.667	16.487
66	3	60 S	60 S	40 S	50 S	52.50	72.66	2.46	0.211	87.496
67	4	90 S	70 S	30 S	40 S	57.50	79.58	1.83	0.828	876.099
68	4	50 S	60 S	10 MR	40 S	38.50	53.29	4.20	0.350	680.265
69	-	70 S	70 S	20 S	20 S	45.00	62.28	3.39	1.102	412.808
70	-	90 S	80 S	40 S	10 Mr	53.50	74.04	2.33	1.840	578.325
71	2	90 S	70 S	30 S	10 Mr	48.50	67.12	2.95	1.669	846.222

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Cont. Table (3)

2011	. I able	(9)								
						plant rea	action			
No.	Seedling		n / Leaf	rust seve	rity %	ACI	CARPA	RRI	b	S²d
		Gemmeiza	Sakha	Nubaria	Ismaylia					
72	4	40 S	60 S	40 S	5 Mr	35.50	49.13	4.57	1.203	9.231
73	-	30 Ms	70 S	50 S	30 S	43.50	60.20	3.58	0.607	413.887
74	3	60 S	60 S	40 S	10 Mr	41.00	56.74	3.89	1.283	55.935
75	-	90 S	80 S	40 S	30 S	60.00	83.04	1.52	1.228	528.803
76	2	30 Ms	70 S	30 S	15 Mr	32.50	44.98	4.95	1.163	271.573
77	3	80 S	70 S	30 S	20 S	50.00	69.20	2.77	1.170	378.263
78	0	40 S	70 S	40 S	10 Ms	39.50	54.67	4.07	1.251	42.193
79	2	90 S	80 S	30 S	20 s	55.00	76.12	2.15	1.448	778.518
80	3	70 S	80 S	70 S	10 Mr	56.00	77.51	2.02	2.015	29.607
81	1	90 S	90 S	30 S	15 Mr	54.00	74.74	2.27	2.117	677.513
82	4	90 S	90 S	70 S	40 S	72.50	100.00	0.00	1.328	70.131
83	4	40 S	60 S	40 S	15 Mr	36.50	50.34	4.46	0.292	634.586
84	3	30 S	70 S	30 S	40 S	42.50	58.62	3.72	0.419	347.090
85	-	40 S	70 S	30 S	10 Mr	36.00	49.65	4.53	1.318	64.716
86	0	60 S	60 S	40 S	15 Ms	41.50	57.24	3.84	1.129	49.772
87	0	60 S	50 S	40 S	30 S	47.50	65.51	3.10	0.508	73.662
88	2	30 Ms	15Mr	20 Ms	10 Mr	12.50	17.24	7.45	0.133	62.301
89	1	40 S	60 S	40 S	20 S	40.00	55.17	4.03	0.783	20.748
90	0	50 S	50 S	30 S	40 S	42.50	58.62	3.72	0.211	87.496
91	0	40 S	60 S	30 S	10 Mr	33.50	46.20	4.84	1.156	15.673
92	3	40 S	70 S	40 S	30 S	45.00	62.06	3.41	0.715	110.741
93	3	30 Ms	50 S	40 S	10 Mr	29.50	40.68	5.33	0.886	138.48
94	0	30 Ms	80 S	40 S	30 S	43.50	60.00	3.60	0.761	553.642
95	2	60 S	70 S	40 S	30 S	50.00	68.96	2.79	0.832	62.352
96	3	70 S	70 S	40 S	20 S	50.00	68.96	2.79	1.120	110.11
97	3	40 S	70 S	60 S	40 S	52.50	72.41	2.48	0.504	146.624
98	3	90 S	80 S	50 S	40 S	65.00	89.65	0.93	1.007	420.259
99	3	70 S	60 S	40 S	30 S	50.00	68.96	2.79	0.728	142.025
100	1	70 S	70 S	40 S	5 Mr	45.50	62.75	3.35	1.540	107.643
101	0	30 Ms	30 Ms	20 Ms	10 Mr	17.00	23.44	6.89	1.187	376.221
102	1	30 Ms	90 S	50 S	10 Mr	42.00	57.93	3.78	1.706	754.424
103	0	60 S	70 S	10 Mr	15 Mr	35.00	48.27	4.65	1.373	589.975
104	0	40 S	50 S	20 S	10 Mr	28.50	39.31	5.46	0.985	47.194
105	3	80 S	70 S	80 S	20 S	62.50	86.20	1.24	1.213	276.174
106	3	60 S	70 S	60 S	30 S	55.00	75.86	2.17	0.850	7.320
107	3	30 S	50 S	60 S	40 S	45.00	62.06	3.41	0.121	193.428
108	3	30 Ms	40 S	80 S	10 Mr	37.00	51.03	4.40	0.743	1131.124
109	0	70 S	70 S	70 S	30 S	52.50	72.41	2.48	0.917	70.456
110	1	90 S	80 S	80 S	15 Mr	64.00	88.27	1.05	1.836	473.381
111	2	40 S	40 S	40 S	10 Mr	31.00	42.75	5.15	0.840	58.911
112	4	30 S	70 S	60 S	30 S	47.50	65.51	3.10	0.675	290.526
113	3	40 S	70 S	70 S	20 S	50.00	68.96	2.79	0.971	263.136
114	3	50 S	70 S	60 S	5 Mr	45.50	62.75	3.35	1.441	91.089
115	-	50 S	90 S	30 S	10 Mr	43.50	60.00	3.60	1.863	319.270
116	4	60 S	70 S	60 S	5 Mr	48.00	66.20	3.04	1.500	73.846
117	2	30 Ms	70 S	60 S	20 S	43.50	60.00	3.60	0.846	486.692
118	3	80 S	50 S	60 S	5 Mr	48.00	66.20	3.04	1.292	521.120
119	1	50 S	40 S	70 S	10 Mr	41.00	56.55	3.91	0.925	525.325
120	4	60 S	50 S	70 S	10 Mr	46.00	63.45	3.29	1.146	408.680

(-)absent General mean : ACI = 37.47 b = 1.00 $S^2d = 239.042$

Accordingly, the genotype No.29, have the highest level of resistance to leaf rust and could be considered as a good source of resistance, while the genotype No.88, 28 and 12 could be scored as reasonable resistant lines. It could be noticed that the lines No. 9, 13 and 101 showed ACI values up to 17 (20.00 , 20.00, 17.00, as well as RRI values 6.51, 6.51 and 6.89 ,respectively). These lines could be improved through crossing with other resistant lines. Dubin and Rajaram (1981) reported that low average of coefficient of infection indicated the presence of broadly-based resistance. Similar results were obtained by Ghobrial *et al.* (1984); Hussain (1997); Rizk *et al.* (1997) and Akhtar, *et al.* (2002) on other barley genotypes.

On the other hand, the desirable / acceptable relative resistance index (RRI) were assessed. Data presented in (Table 4) showed that the desirable barley germplasms with relative resistance index (RRI 7 and above) to leaf rust during 2005 / 2006 season are as follows:

A-Yield trial No. 12, 28 and 29.

B-Yield trial No. 88

Also, the acceptable barley germplasms with relative resistance index (RRI 6 or 5) are as follows:

A-Yield trial No. 5, 9, 10, 13, 14, 15, 16, 20, 21, 26, 27, 41, 44 and 49.

B-Yield trial No. 93

D- Yield trial No. 101, 104 and 111

Similar results which were obtained on other barley genotypes by Akhtar *et al.* (2002) supported this study on barley leaf rust disease.

Table (4): Barley genotypes with desirable/acceptable relative resistance index (RRI) against leaf rust during 2005 / 2006 season.

3 c a3011.		
Genotype No.	Desirable (RRI 7 and above)	Acceptable (RRI 6 or 5)
5	-	5.03
9	-	6.51
10	-	5.46
12	7.26	-
13	-	6.51
14	-	5.89
15	-	5.89
16	-	5.64
20	-	6.27
21	-	5.83
26	-	5.64
27	-	5.34
28	7.32	-
29	8.38	-
41	-	6.02
44	-	5.64
49	-	5.13
88	7.45	-
93	-	5.33
101	-	6.89
104	-	5.46
111	-	5.15

The stability of these one hundred and twenty genotypes against barley leaf rust were evaluated by calculating the stability statistics namely (b) which refer to the regression coefficient of the performance of each of the genotypes under different environments and (S ²d) which refer to the mean square deviation from linear regression were calculated.

The ideal genotype must be characterized by the following characteristics:

- 1- Regression coefficient should be significantly different from zero (b \neq 0) and not significantly different from unity (b = 1).
- 2- Minimum value of the deviation from linear regression S 2 d = 0.
- 3- Low disease severity within a reasonable range of environmental variations.

According to the previous criteria, data in (Table 5) reveal that nine genotypes i.e. (9, 20, 21, 26, 27, 29, 44, 93 & 104) showed the highest stability for resistance to barley leaf rust.

Table (5): Selective barley genotypes expressed by average coefficient of infection (ACI) and stability parameters for resistance to leaf rust disease.

Genotype No.	ACI	b	S ² d
9	20.00	0.514	78.527
20	22.00	0.873	143.003
21	25.50	0.787	8.623
26	27.00	0.724	121.750
27	29.50	0.908	27.716
29	5.00	0.604	20.976
44	27.00	1.030	181.243
93	29.50	0.886	138.48
104	28.50	0.985	47.194

Finally, it can be concluded that:

The genotype No. 29 followed by lines No. 88, 28 & 12 have the highest level of resistance to barley leaf rust and could be considered as a good source of resistance. The lines which showed desirable / acceptable relative resistance index (RRI) in this study are sufficient to be used as parents in breeding programs for developing new disease resistant cultivars.

REFERENCES

Akhtar, M.A.; Rattu,A.R.; Mirza, J.I.; Ahmed, S.J.Hasid, Haque, S.A.J., Hakro, A.A., and Jaffery,A.H.(2001). Evaluation of candidate lines against strip and leaf rust under national uniform wheat and barley yield trial,1999-2000. Pak.J. Phytopathol.,12:45-57.

Akhtar, M.A.; Ahmed, I.; Mirza, J.I. Rattu, A.R.; Haque, E.UI; Hakro, A.A. and Jaffery, A.H.(2002). Evaluation of Candidate Lines Against Stripe and Leaf Rusts Under National Uniform Wheat and Barley Yield Trial,2000-2001. Asian Journal of Plant Sciences, 1(4):450-453.

- Aslam, M.(1982). Uniform procedure for development and release of improved wheat varieties. Mimeograph, PARC, Islamabad.
- Clifford, B . C. ,(1985) . Barley leaf rust. In: A. P. Roelfs, and W. R . Bushnell (eds), The Cereal Rusts , Vol. II, 173 205 , Academic Press, Orlando.
- Czembor JH, Bladenopoulos K, (2007). Resistance to leaf rust (*Puccinia hordei*) in Greek barley cultivars and breeding lines. Cereal Rusts and Powdery Mildews Bulletin www.crpmb.org/2007 / 0215czembor
- Dubin, H.J. and Rajaram,S.(1981). The strategy of the International Maize and Wheat Improvement Center (CIMMYT) for Breeding Disease Resistant Wheat: An International Approach. Strategies for the Control of Cereal Diseases.J.F.Jenkyn and R.T. Plumb (eds.)Federal of British Plant Pathologists. Pp. 27 -35.
- Eberhart , S.A. and Russell, W.A. (1966). Stability parameters for comparing varieties , Crop Sci., : 36 40.
- El Marakby , A. M . ,. Abou Alam, A. M and El Hariry, S.H.M. (1986). Genotypic Stability analysis for some *G. barbadense and G. hirsutum* genotypes of cotton. Ann. Agric. Sci., Moshtohor 24,3: 1347 1367.
- El Sayed , A. A. , Faten El Nashar , Masarat El Ghamry , E. E. Mostafa and Rizk, R. A.,(1991). Disease resistance of barley genotypes under the conditions of Northwestern coast of Egypt . Assiut J . of Agric. Science , Vol. 22 (1) : 127 142 .
- Finaly, K.W. and Wilkinson, G.N. (1963). The analysis of adaptationin plant breeding program. Aust. J. Agric. Res.,14: 742 -754.
- Ghobrial, E.; Rizk, R.A.; Mostafa, E.E. and Abdel-Azim, N.Z.(1984). Sources of multible-disease resistance to barley foliar diseases under Egyptian conditions. Proc. Of the 6th Congr. Of the Un. Phytopath. Mediterr., Cairo, Egypt, 335 337.
- Griffey CA., Das MK, Baldwin RE, Waldenmaier CM, (1994). Yield losses in winter barley resulting from a new race of *Puccinia hordei* in North America. Plant Disease 78, 256-260.
- Hussain ,M.(1997). Report on evaluation of candidate lines against stripe and leaf rusts under National Uniform Wheat ,Barley and Triticale Yield Trials, 1996-97, CDRI, NARC Pak. Agric. Res. Conc., pp:23.
- Martinez, F.; Niks, R.E.; Moral, A.; Urbano, J.M.; Rubiales, D.and Hernande, P. (2001). Search for partial resistance to leaf rust in a collection of ancient Spanish wheat. Proceeding of the fourth International Triticeae Symposium, Cordoba, Spain. September 2001 Hereditas Lund 2001, 135: 193 197.
- Mirza, J.I.; Singh, R.P.; and Ahmed, I. (2000). Resistance to leaf rust in Pakistan wheat lines. Pak. J. Biol. Sci., 3:1056-1061.
- Nabila, A. Mostafa, and. Ahmed, I. A. (2005). Assessment of new genetic resources for controlling powdery mildew and leaf rust diseases of barley. Egypt. J. of Appl. Sci., 20 (10 p).
- Niks RE, Walther U, Jaiser H, Martinez F, Rubiales D, Andersen O, Flath K, Gymer P, Heinrichs F, Jonsson R, Kuntze L, Rasmussen M, Richter E, (2000). Resistance against barley leaf rust (*Puccinia hordei*) in West European spring barley germplasm. Agronomie 20, 769 782.

- Nooman , M.M., Rizk, R.A., Massarat El Ghamry, Faten El Nashar , E. E. Mostafa and Asaad, F.A. (1992). Barley genotypes as influenced by major diseases under irrigated and rain fed conditions. Zagazig J. Agric. Res. Vol. 19 (1): 211 225.
- Park RF, (2003). Pathogenic specialization and phenotype distribution of *Puccinia hordei Otth.* in Australia , 1992 2001. Plant Disease 87, 1311 1316.
- Peterson, R.F.; Campell, A.B.; and Hanna, A,E. (1948). A diagrammatic scale for estimating rust intensity on leaves and stems of cereals. Can. J.Res. C., 26: 496 500.
- Prescott, J. M. and Saari, E.E. (1975). Major disease problems of durum wheat and their distribution within the region. 3rd Regional wheat workshop, Tunis, Tunisia. 104 114, April 28 May 2.
- Rizk, R. A.; M. A. Meghad; E. E. Mostafa and Faten EI Nashar (1997). Evaluation of some barley genotypes for disease resistance and yield potential under different irrigation systems. Egypt. J. Appl. Sci., 12 (12): 742 750.
- Saari, E.E. and Wilcoxson, R.D. (1974). Plant disease situation of high yielding durum wheat in Asia and Africa. Annual Review of Phytopathology, 73: 931 935.
- Snedecor, G.W. and Cochran, W.G. (1967). Statistical Methods. 6th . Iowa State Univ. Press, Iowa , USA.
- Stakman, E.C.; Stewart, D.M.; and Loegering, W.Q. (1962). Identification of physiologic races of Puccinia graminis var. tritici. U.S.D.A., Agric. Res. Serv. Bull. E 617.
- Tarvet,I.. and Cassell, R.C. (1951). The use of cyclone separation in race identification of cereal rust. Phytopathology, 41: 282.
- Whelan, HG: Gaunt, R.E.: Scott WR, (1997). The effect of leaf (*Puccinia hordei*) on yield response in barley (Hordeum vulgare L.) crops with different yield potentials. Plant Pathology, 46: 397 406.

مقاومة بعض التراكيب الوراثية لمرض صدأ الأوراق فى الشعير المتسبب عن الفطر بكسينيا هورديياي

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تم تقييم ١٢٠ تركيب وراثى (سلالة) من الشعير ضد مرض صدأ الأوراق نمثل اربع تجارب محصولية وهي (أ – ب – ج – د) و ذلك في طور البادرة تحت ظروف العدوي الصناعية بالصوبة و طور النبات البالغ في أربعة مواقع خلال موسم ٢٠٠٥ - ٢٠٠٦ . اظهر عدد ٥٥ تركيب وراثي درجة من المقاومة (طراز إصابة منخفض) في طور البادرة تمثل ٤٥,٨٣ % من إجمالي التراكيب المختبرة و ذلك مقارنة بالأصناف التجارية الثلاثة القابلة للإصابة في كل مجموعة (طراز الإصابة مرتفع) . من بين هذه التراكيب الوراثية وجد ٢٠ سلالة عالية المقاومة حيث اظهرت طراز إصابة منخفض . ايضاً تم تقييم هذه التراكيب الوراثية في أربعة مواقع متباينة في الظروف البيئية وهي سخا - الجميزة - النوبارية - و الإسماعيلية حيث تم حساب شدة الإصابة و متوسط معدل الإصابة و المقياس النسبي للمقاومة بالإضافة إلى انـه امكن تحديد التراكيب الوراثية المرغوبة و المقبولة بالنسبة للمقاومة لمرض صدأ الأوراق . من خلال التحليل الإحصائي بإستخدام معامل الإنحدار للتراكيب الوراثية المستخدمة تحت الظروف المتباينة تم حساب درجة ثباتها للمقاومة . اظهرت معظم التراكيب الوراثية درجة من القابلية للإصابة بمرض الصدأ و كان متوسط شدة الإصابة بالإسماعيلية أقل منها من المواقع الاخرى و تبين أن سلالة الشعير رقم ٢٩ من أفضل السلالات من حيث المقاومة و الثبات الوراثي على مستوى المواقع الأربعة حيث كان متوسط معدل الإصابة (٥,٠٠) و المقياس النسبي للمقاومة (٨,٣٨) يليها السلالات رقم ٨٨ ، ٢٨ ، ١٢ من حيث المقاومة حيث وجد أن متوسط معدل الإصابة بين (١٢,٥ - ١٤,٠٠) و المقياس النسبي للمقاومة (٧,٤٥ - ٧,٢٦). و هذه التراكيب الوراثية يمكن استخدامها كأباء في برنامج تربية أصناف جديدة من الشعير مقاومة لمرض صدأ الأوراق.