

Journal of Plant Protection and Pathology

Journal homepage & Available online at: www.jppp.journals.ekb.eg

Sensitivity of *Botrytis cinerea* Isolates Collected from Strawberry to SDHI Fungicide Boscalid

Al-Zahraa E. Ahmed¹; Y. M. Shabana² and M. S. Hamada^{1*}



Cross Mark

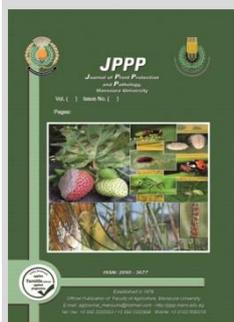
¹Pesticides Department, Faculty of Agriculture, Mansoura University, Mansoura 35516, Egypt

²Plant Pathology Department, Faculty of Agriculture, Mansoura University, Mansoura 35516, Egypt

ABSTRACT

Egypt is considered one of the main strawberry producers countries. Grey mould, caused by *Botrytis cinerea*, is a destructive disease resulting in significant losses in strawberry production worldwide as fruits might be infected in the field, storage and transport. Management approaches for the pathogen in strawberries depend mainly on using protective fungicides. Recently, lack of boscalid fungicide efficiency reported in many countries. The current study aims to detect and monitor *B. cinerea* resistance to boscalid in the main strawberry production governorates in Egypt (Beheira, Ismailia and Qalyubie) between 2019 and 2021. 269 isolates were collected and tested to distinguish resistant isolates. Sensitivity of a set of resistant and sensitive isolates was measured by determining the effective concentration that inhibits 50 % of mycelial growth. The results showed that, 225 (83.6%) isolates were resistant to boscalid while, only 44 (16.4%) isolates were sensitive. Monitoring of resistance frequency revealed that there were remarkable increases in resistance frequencies to boscalid in *B. cinerea* isolates during 2019 and 2021 seasons as it was 84.03% in 2019 increased to 98.87% in 2021. 17 sensitive isolates randomly selected were used to determine the EC₅₀ for boscalid using mycelial growth inhibition assay and the EC₅₀ values ranged from 0.0067 to 1.14 µg/ml with a mean of 0.29 µg/ml. While, the EC₅₀ values for 17 resistant isolates randomly selected ranged from 2.1 to 16.2 µg/ml with a mean of 7.3 µg/ml. Among 225 boscalid resistant isolates, 192 (85.33%) isolates showed resistant to pyraclostrobin which may lead to failure of gray mold control in case of usage of those fungicides mixture in control strategies.

Keywords: *Botrytis cinerea*, Strawberry, SDHI, Boscalid, Resistance



INTRODUCTION

Strawberry is an important horticultural crop, widely grown in all temperate regions in the world. Gray mold, caused by *Botrytis cinerea* (*B. cinerea*), is among the most important diseases of several crops as it can infect over than 200 plant species including strawberry, causing significant yield losses worldwide. *B. cinerea* found in the remains of the botanical parts. High humidity helps develop disease. With the start of flowering, fungi attack flowers, which lead to less flowers and loss of fruits (Adnan *et al* 2019).

Control of gray mold in strawberry fields mainly depends on usage of protective fungicides extensively in order to avoid the infection starting process. Recently, growers are facing a real challenge due to less efficiency of common fungicides used as a result of rapid development of *B. cinerea* resistant populations (Amiri *et al* 2013). Resistance to multiple chemical classes of fungicides, such as Quinone outside inhibitors (QoIs), Hydroxylanilides (HAs) or Phenylpyrroles (PPs) reported in many studies (Bardas *et al* 2010 and Fan *et al* 2017) complicated the control process and lead to failure in the strategies used to minimizing the losses. Succinate dehydrogenase inhibitors (SDHIs) also known as succinate: quinone oxidoreductase or succinate dehydrogenase such as boscalid is wide spectra fungicides used recently in *B. cinerea* control. According to FRAC SDHI fungicides are classified as medium to high risk of resistance development. Boscalid was the first molecule belonging to the SDHIs that was introduced for the control of

B. cinerea. The target of the SDHI fungicides is inhibiting complex II in the mitochondrial respiratory chain, leading finally to blocking the cell energy production. *B. cinerea* boscalid resistant populations were detected in many countries worldwide. Wang *et al* (2021) mentioned that 45.16% of *B. cinerea* isolates were resistant to boscalid. Similarly, Cosseboom *et al* (2019) stated that 24% of isolates were resistant to the fungicide. Additionally, monitoring the resistance frequency is very important in order to prolong the effective use of the fungicide. Cui *et al* (2021) tested the sensitivity of *B. cinerea* isolates to boscalid and the results illustrated that the resistance frequency increased from 5.26% and 0 to 56.25% and 52% for Laiwu and Liaocheng locations respectively, after five years of intensive usage of this fungicide. Resistance to boscalid linked to several mutations in one of the subunits (B, C, or D) of the SDH complex (Hu *et al* 2016).

Using of fungicides combinations is an effective strategy to delay the development of resistant populations. Boscalid and quinone outside inhibitor (QoI) fungicide pyraclostrobin mixture used recently to control *B. cinerea* in Egypt widely. Worldwide Pristine® is a combination of the SDHI and QoI fungicides used in the control of gray mold (kim and Xiao 2010 and Fernández-Ortuño *et al* 2012).

Therefore, the objectives of the current study were to detect boscalid resistance frequencies in *B. cinerea* isolates collected from different governorates in Egypt, monitor resistance to SDHI fungicide in *B. cinerea* from strawberry

* Corresponding author.

E-mail address: m_sobhy@mans.edu.eg

DOI: 10.21608/jppp.2022.161968.1093

fields and to determine the effective concentrations EC_{50} of the sensitive and resistant isolates. Moreover, to investigate the dual resistance between boscalid and pyraclostrobin.

MATERIALS AND METHODS

Fungal isolates and culture conditions:-

In this study, 269 *B. cinerea* isolates were collected between 2019 and 2021. The governorates selected for isolates collection were Beheira, Ismailia and Qalyubie. Each governorate 4 commercial annual strawberry fields were chosen. Strawberry fruit carrying typical symptoms of gray mold obtained from commercial fields; one fruit picked from a different plant. Isolates of *B. cinerea* were cultured on potato dextrose agar medium (200 ml of potato juice originated from 200 g of potato, 20 g of agar and 20 g of dextrose, distilled water to 1 L) at 20°C in darkness. Each year, the isolation and purification of the fungal strains was done using a single hyphal tip method. The isolates slants were preserved in 15-mL plastic tubes containing PDA medium at 4°C until use.

Active ingredients: -

Technical grade boscalid (96% a.i.; Zhejiang Heben Pesticide & Chemicals Co., Ltd). Technical grade pyraclostrobin (98% a.i.; Shanghai Heben-Eastsun Medicaments Co., Ltd.) The fungicides dissolved in 100% acetone to produce a stock solution containing 100 mg/ml.

According to previous studies, Hu *et al* (2016), Chen *et al* (2016) and Fan *et al* (2017) 75 µg/ml was used as a discriminatory concentration (a concentration that fully inhibits mycelial growth of the sensitive isolates) to distinguish boscalid resistant isolates from sensitive isolates. 5 mm in diameter mycelial plugs were separated with a cork borer from 3 day old colony margin of each isolate and placed upside down onto the media amended with fungicide. Colony diameter measured after 3 days at 22°C. For each fungicide, the isolates considered resistant if they can grow on the media amended with fungicide, while isolates cannot grow on the amended media were considered to be sensitive isolates.

Sensitivity of *B. cinerea* to boscalid:-

The sensitivity to boscalid was determined by evaluating the effective concentration resulting in inhibition of 50% of mycelial growth (EC_{50}) values for 17 boscalid sensitive (Bos^S) and 17 boscalid resistant (Bos^R) isolates. Technical grade boscalid (96% active ingredient [a.i.] was dissolved in 100% acetone, adjusted to a concentration of 100 mg/ml, and added to PDA to produce final concentrations at (0, 0.1, 1, 2, 4 and 8 µg/ml) and (0, 10, 25, 50, 75 and 100 µg/ml) to test sensitive isolates and resistant isolates, respectively. The experiment was repeated twice. Data Processing System (DPS) program developed by Hangzhou Reifeng Information Technology Ltd., Hangzhou, China used to calculate the EC_{50} value for each isolate. The average of EC_{50} values from the two experiments for each isolate used in the analysis of the data as there was no significant difference ($P>0.05$) between the two experiments (Hamada *et al* 2011).

Dual resistance between pyraclostrobin and boscalid

To screen dual resistance in *B. cinerea* isolates between pyraclostrobin and boscalid, all boscalid resistant isolates were tested for sensitivity to pyraclostrobin. Discriminatory concentration of 10 µg/ml used to detect Pyraclostrobin resistant isolates also salicyl hydroxamic acid (99% a.i.; Sigma-Aldrich, St. Louis) at 100 µg/ml added to

potato dextrose agar (PDA) to inhibit the alternative oxidase respiration.

RESULTS AND DISCUSSION

Resistance in *B. cinerea* isolates to SDHI fungicide boscalid

Among 269 isolates were collected, 225 (83.6%) isolates were resistant to boscalid fungicide as they were able to grow in media amended with discriminatory concentrations 75 ppm while, only 44 (16.4%) isolates were sensitive due to high selection pressure occurred. Moreover, it was observed that the percentage of resistance to boscalid was 84.03% in 2019 while it decreased in 2020 to 68% and again increased in 2021 to 98.87%.

In harmony with our results, high resistance frequencies reported by many authors such as Fernandez *et al.*, 2017 in Spain who mentioned that the percentage of resistance to boscalid in *B. cinerea* isolates collected from strawberries recorded was 73%. Also, Chen *et al* (2016) found that 66.67% of *B.cinerea* isolates collected were resistant to boscalid. While moderate resistance frequencies were reported by Weber *et al.*, 2011 in Northern German who reported that 21.5% of isolates were resistant to boscalid in the isolates from strawberry. On the other hand, Yin *et al.*, 2018 stated that the percentage of resistance to boscalid was only 4.3% among the isolates collected from nectarine and cherry in China. According to our results, there was remarkable increase in resistance frequencies to boscalid in *B. cinerea* isolates during 2019 and 2021 seasons as it was 84.03% in 2019 and increased to 98.87% in 2021. However, the resistance frequency decreased in 2020 to 68% which could be returned to the decrease in selection pressure occurred as a result of usage of different fungicides in this season or due to climatic changes. Many authors monitored the increasing and decreasing of the percentage of resistance in *B. cinerea* isolate to boscalid such as Rupp *et al.*, 2016 in Germany who observed that the percentage of resistance to boscalid remarkably increased between 2001 to 2014 from 4% to 35%. In the same way, Leroch *et al.*, 2011 found that frequencies of resistance to boscalid increased gradually among *B. cinerea* isolates collected from grape during the study as it was 2% in 2006 while it 26.7% in 2009. Also, Fernández-Ortuño *et al.*, 2014 observed that percentage of resistance to boscalid was 29% in 2013 while it was 5% in 2012.

Table 1. Information about the sensitivity of 269 *Botrytis cinerea* isolates collected from strawberry in Egypt during three years from 2019 to 2021 to boscalid

| Location | Year of collection | No. of isolates | No. of Bos^S | No. of Bos^R | % of resistance |
|----------------------------|--------------------|----------------------------|----------------|----------------|-----------------|
| Beheira | 2019 | 37 | 3 | 34 | 91.9 |
| Ismailia | 2019 | 43 | 13 | 30 | 69.8 |
| Qalyubie | 2019 | 39 | 3 | 36 | 92.3 |
| Beheira | 2020 | 26 | 4 | 22 | 84.6 |
| Ismailia | 2020 | 26 | 12 | 14 | 53.8 |
| Qalyubie | 2020 | 23 | 8 | 15 | 65.2 |
| Beheira | 2021 | 25 | Zero | 25 | 100 |
| Ismailia | 2021 | 25 | Zero | 25 | 100 |
| Qalyubie | 2021 | 25 | 1 | 24 | 96 |
| Total | / | 269 | 44 | 225 | |
| Resistant percentage% | | | 16.4 | 83.6 | |
| Bos^S boscalid sensitive | | Bos^R boscalid resistant | | | |

Sensitivity of *B. cinerea* isolates to boscalid:-

Among 269 isolates of *B. cinerea* obtained in the study, 17 Bos^S isolates randomly selected were used to determine the EC₅₀ for boscalid using mycelial growth inhibition assays. The EC₅₀ values of boscalid ranged from 0.0067 to 1.14 µg/ml with a mean of 0.29 µg/ml and the highest values EC₅₀ of isolates (52.94%) ranged from 0.1 to 0.9 µg/ml (Fig.1). While, to determine the EC₅₀ values for resistant isolates 17 Bos^R isolates randomly selected and tested using mycelial growth inhibition assays. The EC₅₀ values ranged from 2.1 to 16.2 with a mean of 7.3 µg/ml. and the highest EC₅₀ values of isolates (41.18%) ranged from 1 to 4.9 µg/ml (Fig.1)

RF (Resistance Factor) values to boscalid calculated by dividing the EC₅₀ value of the resistant isolate by the mean EC₅₀ value of the sensitive isolates (Table2) and the results illustrated that the resistant factor ranged between 7.26 to 55.8 which cleared that different levels of resistance could be found.

Table 2. EC₅₀ of *Botrytis cinerea* (sensitive and resistant) isolates randomly selected to Boscalid:

| Boscalid (EC ₅₀) | | | | |
|------------------------------|------------------|-----------|------------------|-------|
| Sensitive | | Resistant | | |
| Isolate | EC ₅₀ | Isolate | EC ₅₀ | RF |
| S 108 | 0.0068 ± 0.0002 | 8 B | 2.1066 ± 0.0004 | 7.26 |
| S 29 | 0.0305 ± 0.0003 | S 3 | 2.5207 ± 0.0099 | 8.69 |
| 39 | 0.0316 ± 0.0002 | BN 80 | 2.7407 ± 0.0003 | 9.45 |
| 13 | 0.0397 ± 0.0002 | H4(1) | 3.2938 ± 0.0002 | 11.36 |
| BN 2 | 0.0728 ± 0.0001 | 36 | 3.9833 ± 0.0006 | 13.74 |
| S 43 | 0.0737 ± 0.0002 | MB 33 | 4.0720 ± 0.0009 | 14.04 |
| S 35 | 0.0812 ± 0.0002 | 1 | 4.7259 ± 0.0013 | 16.29 |
| 19 B | 0.1163 ± 0.0003 | 121 | 6.0356 ± 0.0006 | 20.81 |
| H4(32) | 0.1191 ± 0.0002 | H3(6)2 | 6.1872 ± 0.0006 | 21.34 |
| H4(19) | 0.1313 ± 0.0002 | MB 38 | 7.7608 ± 0.0010 | 26.76 |
| S 137 | 0.1984 ± 0.0002 | S 147 | 7.8016 ± 0.0006 | 26.9 |
| S 28 | 0.2771 ± 0.0003 | 7 | 8.1572 ± 0.0005 | 28.13 |
| 48 | 0.3767 ± 0.0002 | 1 B | 8.9026 ± 0.0009 | 30.69 |
| 21 B | 0.6477 ± 0.0002 | BN 105 | 10.8557 ± 0.0003 | 37.75 |
| S 37 | 0.7085 ± 0.0002 | H4(35) | 14.0823 ± 0.1109 | 48.38 |
| S 159 | 0.9676 ± 0.0002 | H2(12)2 | 15.1577 ± 0.2007 | 51.99 |
| 49 | 1.1397 ± 0.0002 | 132 | 16.2061 ± 0.1599 | 55.8 |

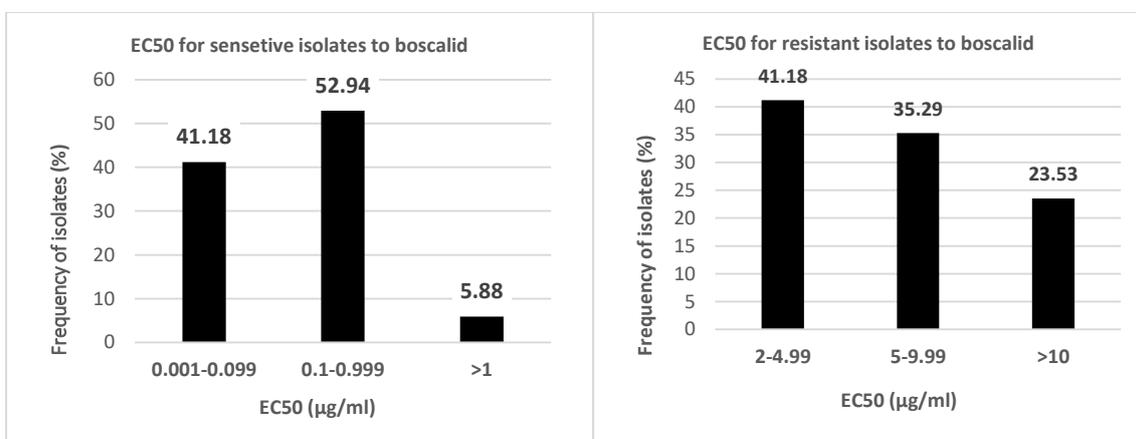


Fig 1. Sensitivity of *B. cinerea* isolates to boscalid

Our results were in harmony with many authors such as Chatzidimopoulos *et al* (2013) they studied the sensitivity of *B. cinerea* to boscalid by the determination of EC₅₀ values and the results showed that the EC₅₀ ranged from 0.38 to 2.7 mg /L using mycelial growth inhibition assay. Also, Pokorny *et al* (2016) in California reported that the effective concentration values of boscalid for *B. cinerea* sensitive isolates were 0.13 and 0.29 ppm while it was more than 100 ppm for resistant isolates.

Resistance to pyraclostrobin and boscalid

The results obtained in the current study indicated that among 225 boscalid resistant isolates detected 192 (85.33%) isolates showed resistance to pyraclostrobin which indicated that using of a mixture of both fungicides may result in the failure of the pathogen control. In harmony with our results, Fernández-Ortuño *et al* (2012) reported that 61.5% of *B. cinerea* isolates tested were resistant to both pyraclostrobin and boscalid. Moreover, kim and Xiao (2010) recorded that 20% of the isolates tested showed resistance to pyraclostrobin and boscalid.

CONCLUSION

The usage of single site fungicides such as boscalid extensively as a part of *B. cinerea* control strategies have to be performed accurately to reduce the risk of resistance in strawberries. According to the results obtained in the current

study, *B. cinerea* resistance to boscalid was recorded widely at all locations tested, and it is recommended to reduce the number of applications of this fungicide in the same season and use it alternating with other fungicides in order to control this disease effectively. The mixture of pyraclostrobin and boscalid should not be the first option in control strategies in Egypt in order to manage boscalid resistant populations.

REFERENCES

Adnan, M., Hamada, M. S., Hahn, M., Li, G. Q., and Luo, C. X. (2019). Fungicide resistance of *Botrytis cinerea* from strawberry to procymidone and zoxamide in Hubei, China. *Phytopathol. Res.* 1:17.

Fan, F., Hamada, M. S., Li, N., Li, G. Q., and Luo, C. X. (2017). Multiple fungicide resistance in *Botrytis cinerea* from greenhouse strawberries in Hubei province, China. *Plant Dis.* 101, 601–606.

Bardas, G. A., Veloukas, T., Koutita, O., and Karaoglaniadis, G. S. (2010). Multiple resistance of *Botrytis cinerea* from kiwifruit to SDHIs, QoIs and fungicides of other chemical groups. *Pest Manage. Sci.* 66:967-973.

Wang, W.; Fang, Y.; Imran, M.; Hu, Z.; Zhang, S.; Huang, Z.; Liu, X. (2021). Characterization of the Field Fludioxonil Resistance and Its Molecular Basis in *Botrytis cinerea* from Shanghai Province in China. *Microorganisms* 9, 266.

- Cosseboom, S.D., Ivors, K.L., Schnabel, G., Bryson, P.K., and Holmes, G.J., (2019). Within-season shift in fungicide resistance profiles of *Botrytis cinerea* in California strawberry fields. *Plant Disease*, 103, 59–64.
- Cui, Kaidi, He, Leiming, Li, Tongtong, Mu, Wei, Liu, Feng (2021). Development of boscalid resistance in *Botrytis cinerea* and an efficient strategy for resistance management. *Plant Dis.* 105: 1042-1047.
- Hu, M.J.; Fernández-Ortuño, D.; Schnabel, G. (2016). Monitoring resistance to SDHI fungicides in *Botrytis cinerea* from strawberry fields. *Plant Dis.* 100, 959–965.
- Chen, S. N., Luo, C. X., Hu, M. J., and Schnabel, G. (2016). Fitness and competitive ability of *Botrytis cinerea* isolates with resistance to multiple chemical classes of fungicides. *Phytopathology* 106:997-1005.
- Hamada, M.; Yin, Y. and Zhonghua, M. (2011) Sensitivity to iprodione, difenoconazole and fludioxonil of *Rhizoctonia cerealis* isolates collected from wheat in China. *Crop Protec.*, 30: 1028-1033.
- Fernandez-Ortuno, D., Perez-Garcia, A., Chamorro, M., Eduardo, D. L. P., De, Vicente. A., and Tores, J. A. (2017). Resistance to the SDHI fungicides boscalid, fluopyram, fluxapyroxad, and penthiopyrad in *Botrytis cinerea* from commercial strawberry fields in Spain. *Plant Dis.* 101: 1306-1313.
- Weber, R. W. S. (2011). Resistance of *Botrytis cinerea* to multiple fungicides in Northern German small-fruit production. *Plant Dis.* 95:1263-1269.
- Yin, W. X., Adnan, M., Shang, Y., Lin, Y., and Luo, C. X. (2018). Sensitivity of *Botrytis cinerea* from nectarine/cherry in China to six fungicides and characterization of resistant isolates. *Plant Dis.* 102, 578.
- Rupp S, Weber, R.W.S., Rieger, D., Detzel, P. and Hahn, M. (2017). Spread of *Botrytis cinerea* Strains with Multiple Fungicide Resistance in German Horticulture. *Front. Microbiol.* 7:2075.
- Leroch, M., Kretschmer, M., and Hahn, M. (2011). Fungicide resistance phenotypes of *Botrytis cinerea* isolates from commercial vineyards in south West Germany. *J. Phytopathol.* 159:63-65.
- Fernández-Ortuño, D., Grabke, A., Bryson, P. K., Amiri, A., Peres, N. A., and Schnabel, G. (2014). Fungicide resistance profiles in *Botrytis cinerea* from strawberry fields of seven southern U.S. states. *Plant Dis.* 98:825-833.
- Chatzidimopoulos, M., Papaevaggelou, D. and Pappas, A.C. (2013). Detection and characterization of fungicide resistant phenotypes of *Botrytis cinerea* in lettuce crops in Greece. *European Journal of Plant Pathology*, 137, 363–376.
- Pokorny, A.; Smilanick, J.; Xiao, C.-L.; Farrar, J.J.; Shrestha, A. (2016). Determination of fungicide resistance in *Botrytis cinerea* from strawberry in the central coast region of California. *Plant Health Prog.* 17, 30–34.
- Amiri, A., Heath, S. M., and Peres, N. A. (2013). Phenotypic characterization of multifungicide resistance in *Botrytis cinerea* isolates from strawberry fields in Florida. *Plant Dis.* 97:393-401
- Kim, Y. K., and Xiao, C. L. (2010). Resistance to pyraclostrobin and boscalid in populations of *Botrytis cinerea* from stored apples in Washington State. *Plant Dis.* 94:604-612.
- Fernández-Ortuño, D., Chen, F., and Schnabel, G. (2012). Resistance to pyraclostrobin and boscalid in *Botrytis cinerea* isolates from strawberry fields in the Carolinas. *Plant Dis.* 96:1198-1203

حساسية عزلات البوترائيس سناريا التي تم تجميعها من الفراولة لمبيد البوسكاليد
الزهران السيد أحمد¹، ياسر محمد نور الدين شبانة² و محمد صبحي حماده^{1*}
قسم المبيدات- كلية الزراعة - جامعة المنصورة
قسم أمراض النبات - كلية الزراعة - جامعة المنصورة

المخلص

تعتبر مصر من الدول الرئيسية المنتجة للفراولة. العفن الرمادي الذي يسببه فطر بوترايتس سينيريا هو مرض مدمر يؤدي الي خسائر كبيرة في إنتاج الفراولة في جميع أنحاء العالم. قد تصاب الثمار في الحقل والتخزين والنقل. تعتمد أساليب إدارة العوامل الممرضة في الفراولة بشكل أساسي علي استخدام مبيدات الفطريات الوقائية. البوسكاليد هو مبيد فطري جديد واسع الطيف يستخدم للسيطرة علي فطر البوترائيس سينيريا في الفراولة. في الأونة الأخيرة تم ملاحظة نقص كفاءة العديد من المبيدات الفطرية المستخدمة في مكافحة فطر بوترايتس سينيريا في العديد من البلدان. تهدف الدراسة الحالية الي الكشف عن عزلات بوترايتس سينيريا المقاومة للبوسكاليد في محافظات إنتاج الفراولة الرئيسية في مصر (البحيرة، الإسماعيلية، القليوبية) بين عامي 2019، 2021. تم جمع 269 عزلة واختبارها لتمييز العزلات المقاومة والحساسية عن طريق تحديد التركيز الفعال الذي يثبط 50% من نمو الفطريات. أظهرت النتائج أن 225 عزلة (83.6%) كانت مقاومة للبوسكاليد بينما 47 عزلة (16.4%) كانت حساسة. كانت هناك زيادة ملحوظة في ترددات المقاومة للبوسكاليد في عزلات بوترايتس سينيريا. تراوحت قيم التركيز المثبط 50% من نمو العزلات ل 17 عزلة حساسة من 0.0067 الي 1.14 ميكروجرام/ مل بمتوسط 0.29 ميكروجرام/ مل. بينما تراوحت قيم التركيز المثبط 50% من نمو العزلات ل 17 عزلة مقاومة من 2.1 الي 16.2 ميكروجرام/ مل بمتوسط 7.3 ميكروجرام/ مل. بلغ عدد العزلات المقاومة لكلا من مبيد البوسكاليد و مبيد البيروكلستروبين 192 مما يشير الي صعوبة الاعتماد على مخلوط كلا المبيدين في برامج المكافحة