Effect of Some Aromatic Plant Extracts, Silica Nanoparticles and Imidacloprid on the Cotton Whitefly, *Bemisia tabaci* (Genn.) and its Associated Parasitoid, *Eretmocerus mundus* Mercet on Tomato Khafagy, I. F.¹; Amira Sh. M. Ibrahim² and Asmaa M. A. El-Ghobary¹ ¹Plant Protection Research Institute, Sakha Agric. Res. Station, ARC ²Economic Entomology Dept., Fac. Agric., Kafrelsheikh University.



ABSTRACT

This study was conducted at the Experimental Farm belonging to Sakha Agricultural Research Station, Kafr El-Sheikh, ARC, during 2016 and 2017 seasons to investigate the role of aqueous extracts of nine aromatic plants, silica nanoparticle, and the insecticide Imidacloprid (admire) on *Bemisia tabaci* (Genn.) infesting tomato plants . Results showed that the highest value of unhatched *B. tabaci* eggs was 84.40 and 84.60 %respectively on tomato plants treated with the highest concentration of geranium extract, which was significantly different from the other treatments. Silica nano particles killed 90.99 and 91.00 %resp. of early nymphal stage , which was not significantly different from mortality caused by admire . Silica nano particles, spurge and geranium extracts prevented adult development and killed adults at levels that were not significantly different from admire . However, the extracts of four plants; geranium, peppermint, spurge and rosemary had high repellent effect against whitefly adults . The highest parasitism percentage (93.88 and 93.63% respectively) was recorded on tomato plants treated with the highest concentration of sweet basil extract. While the lowest parasitism percentage (16.53 %in both the two seasons) was obtained on tomato plants treated with imidacloprid.

Keywords: *Bemisia tabaci*, silica nanoparticles, tomato, aromatic plants, *Eretmocerus mundus* Corresponding auther: E-mail: khafagyibrahi@yahoo.com

INTRODUCTION

Tomato, *Lycopersicon esculentum* Mill of Family Solanaceae is one of the most consumed vegetables in the world, and global production is estimated at around 136 billion ton per year (FAOSTAT 2009). It is the third most economical important vegetable crop after potato and onion. Tomato is a dietary source of vitamins, especially A and C minerals and fibers which are important for human nutrition and health. Also, tomato is rich in lycopene, a phytochemical that protects cells from oxidants linked to human cancer (Giovannuci 1999 and Mutanen *et al.* 2011). It is also rich in flavonoids and phenolic acid.

The cotton whitefly, Bemisia tabaci (Genn.) (Homoptera: Aleurodidae) is a key pest of vegetables (Al-Musa et al 1987). It is also a serious economic pest of agronomic, horticultural and ornamental crops throughout warm regions of the world (Byrne et al. 1990 and Brown 1994). B. tabaci can develop and reproduce on over 500 plant species, distributed in 74 families (Greathed 1986), and affect some 30 cash and staple crops worldwide, such as tomato, pepper, melon, watermelon, soybean, cotton, beans and cassava. Crop damage may occur directly through excessive sap removal, or indirectly by promoting the growth of sooty mold, inducing syndromes through feeding, or by vectoring plant viruses (Schuster et al. 1996). Estimated economic losses amount to several hundreds or even thousands of millions of dollars a year, worldwide (Oliveira et al. 2001). Begomoviruses are the most numerous of B. tabaci transmitted viruses and can cause crop yield losses between 20 and 100 % (Brown & Bird 1992). B. tabaci is capable of transmitting more than100 different virus species of which the majority belong to the genus geminovirus, such as Tomato Yellow Leaf Curl Virus, Tomato Mottle Virus (Jones 2003) and African Cassava Mosaic Virus (Maruthi et al. 2001). In addition, it has been documented that the action threshold is very low (0.3 adults/plant) in Costa Rica (Hilje 2001).

Plants may provide an alternation to currently used pesticides for control of plant pests, as they constitute a rich source of bioactive chemicals (Kim *et al.* 2005 and Daoubi *et al.* 2005). Recent studies have demonstrated the insecticidal properties of chemicals derived from plants that are active against specific target species, biodegradable to nontoxic products and potentially suitable for use in integrated management programs (Tare *et al.* 2004).

The demand for alternative pest control strategies is increasing. In order to further development, the utilization of semi chemicals or mixing planting strategies is reinvestigated in pest management in polycultures .Volatile chemical compounds emitted from plant tissues are most likely originated to repel the attacking pests, and also serve as a secondary function as it attracts the parasitoids and predators in search of insect pests (Mauchline *et al.*2005).

In the present study, the toxicity of crude extracts of nine aromatic plant species known to have medicinal activity, and silica nanoparticles compared to imidacloprid (Admire) insecticide were investigated against the sweet potato whitefly, *Bemisia. tabaci* and its parasitoid *Eretmocerus mundus* Mercet under semi natural conditions on tomato plants (in wooden frame cage at open field).

MATERIALS AND METHODS

Insect materials:

Adults and immature stages of *Bemisia tabaci* (Genn.) were reproduced on tomato plants for two years .The adults of the parasitoid (*Eretmocerus mundus* Merecet) were collected from different vegetable plants parasitized immature stages of *B. tabaci*. This was practiced by introducing the immature stages of *B. tabaci* in glass- tube cages, till the emergence of the parasitoid or emergence of whitefly adults.

Plant materials:

Tomato (*Lycopersicon esculentum* Mill),Giza 186 variety, was cultivated in pots and the aromatic plants (Table 1) were collected from the local market .

Preparation of the aqueous extracts

The aqueous extracts were prepared by boiling air – dried plant parts in sterile distilled water (10 % wt/wt) for 10 minutes, and then cooled to room temperature overnight . The aqueous extracts were filtered using a Millipore filters (Millipore 0.2 mm, www. Waters. Com) to remove particulate matter. The final volume of each filtrate was completed to 100 ml with distilled water with 0.2% Tween 80 to account for the evaporated water during boiling. The aqueous extracts were prepared shortly before application . Negative controls were represented by the distilled water that contained the emulsifier Tween 80.

 Table 1. Common name, family, scientific name,

 Arabic name and part used of the aromatic

 nlants collected from the local market

plants concercu il olli the local market							
Common	Scientific	Plant	Plant				
Name	name	family	part				
Geranium	Pelargonium graveolens L.	Geraniaceae	Leaves				
Spearmint	Mentha viridis L.	Lamiaceae	Leaves				
Peppermint	Mentha piperita L.	Lamiaceae	Leaves				
Hot-pepper	Capsicum annum L.	Solanaceae	Leaves				
Sweet basil	Ocimum basilicum L	Lamiaceae	Leaves				
Christ's	Ramnus diffusa L.	Rhamnaceae	Leaves				
Rosemary	Rosmarinus officinalis	Lamiaceae	Leaves				
Mooring	Lagopus scoticus L.	Phasianidae	Leaves				
Spurge	Euphorbia hierosolymitana Boiss	Euphorbiaceae	Stems				

Treatments:

Crude extracts of the nine aromatic plants were each applied at the rates of 50, 100 and 150 ml /100 liter of water

Silica nanoparticles: The silica nanoparticles were obtained from Egypt Nanotech Company Limited , Cairo, Egypt. The size of silica nanoparticles was 100 nm with a purity of 99.99 % .It is was applied at the rates of 100, 200 and 300 ppm /100 liter of water

Insecticide: The tested insecticide used in this study was imidacloprid (35% SC) produced by Shreeji Pesticides at the rate of 300 ml/ Feddan. This insecticide is recommended for control *B. tabaci* in vegetable crops (Walaa,2013)

Equipment:

Stereomicroscope: to examine insect stages and their parasitoids.

Aspirators: for collecting and transferring adult insects and parasitoids.

Solo motor, 2.5 Liters volume, a sprayer used in applying the crude extracts of the nine aromatic plant species, silica nanoparticle and Admire insecticide.

A blender, V 2.5 liters for preparing the crude extracts of the aromatic plants .

Paper bags for transferring the plant samples to the laboratory

Cages:

Glass- tube cages: (3 cm in diameter and 20 cm long), for transferring adults of *B. tabaci* to infest tomato plants and to release the parasitoid *E. mundus* Mercet on *B. tabaci* nymphs.

Chimney – **glass cages**: (No. 7) for biological experiments to infest tomato plants with *B. tabaci* adults and to release the parasitoid.

Wooden frame cages $(250 \times 250 \times 200 \text{ cm})$, covered with double layers of muslin on two sides and glass on the top and both front and back sides . The insect – free plants were selected to be infested with *B. tabaci* adults and release the parasitoid .

A Blackman box (Blackman 1971).

Plastic jar (20 cm diameter, 30 cm high).

Curative effect of crude extracts of nine aromatic plants, silica nanoparticles and insecticide (Admire) on *Bemisia tabaci* :

This experiment was carried out under semi natural conditions (in wooden frame cage at open field) of Sakha Agricultural Research Station (SARC), at kafr El- Sheikh during September, October and November, 2016 and 2017. Crude extracts of the nine aromatic plant species at three concentrations (50, 100 and 150 cm) compared to silica nanoparticles, at three concentrations (100, 200 and 300 ppm) and Admire insecticide were evaluated against B. tabaci stages on tomato plants. Seeds of tomato were germinated in tray .The seedlings were transplanted in pots when aged five leaves (one seedling/pot). Each plant was placed under chimney glass cage (No. 7). Recently emerging B. tabaci adults were released (3 pairs /leaf) on tomato plants (5 plants) with five replicates of each treatment, for about 24 hours . Adults of B. tabaci were aspirated from the plants and the plants were placed in separate wooden cages. The synchronously- developing, uniformly- aged whitefly population were then held until they developed to the appropriate stage

Unhatching egg percentage :

Immediately, after the adult whiteflies were aspirated from the plants, the plants were sprayed with abovementioned treatments. Initial number of eggs prior to application ranged from 300 to 350 per plant. There were five replicates (5 Plants/ replicate) per treatment. Four days after treatment, the unhatching eggs and newly emerging nymphs were counted and the hatchability was calculated.

Early stages mortality percentage:

Five days, after infestation, when first nymphal instar emerged and attached to the leaf, the plants were sprayed as before . There were five replicates (5 Plants/ replicate) per treatment. Four days after treatment, the number of dead nymphs were counted under dissecting microscope. A nymph was considered dead if it was shrunken or its color changed. Normally developed nymphs to adult stage were also counted and their percentages were calculated .

Late pupal stage mortality percentage:

Eight days after infestation, when most nymphs were in the red-eye stage, the plants were sprayed as before. There were five replicates (5 plants/ replicated). Seven days after treatment, when most of the pupae emerged from control plants, the number of empty pupae cases and pupae that failed to emerge were counted and percentage of adult emergence was calculated.

Adult mortality percentage :

A fully expanded leaf was placed in wet moss inside a Blackman box (Blackman 1971). The leaf was dipped into the solution of crude extract treatments and left overnight. About 30 adults were then introduced inside each box. Distilled water was used as a negative treatment and imidacloprid was used as a positive one. The number of dead whitefly adults was recorded after 48 hr. A whitefly adult was considered dead if it was motionless after probing with a camel hair brush. Five replicates were made for each treatment.

Repellency test :

Two fully expanded leaves of tomato were placed individually in vials containing water . One leaf was dipped in one of test materials and the other one was dipped in distilled water . The vials were placed in a plastic jar (20 cm diameter, 30 cm high) covered with fine netting material . About 50 mobilized adults were placed between the two vials .Numbers of adults attracted to each leaf was recorded after 3hr and 24hr . Five replicates were made for each treatment .

Effect of tested materials on the parasitoid *Eretmocerus mundus* under semi- natural conditions:

The experiment was carried out under seminatural conditions (in wooden frame cage at open field) during September, October and November 2016 and 2017 seasons, in order to evaluate the efficacy of the nine crude extracts, silica nanoparticles and insecticide at commercial dose on the parasitoid (E. mundus) of B. tabaci on tomato plants. Tomato plants were prepared as previously mentioned . These plants were divided into 32 groups; each group consisted of five plants in four replicates. These tomato plants were placed under chimney-glass cages as one plant/ cage, and recently emerging B. tabaci adults were released as three pairs / leaf, and left for 24 hours. B. tabaci adults were discarded by aspirator. Eight days later, to get the 2ⁿ nymphal instar and preferred the parasitoid according to Jones and Greenberg, 1998 respectively.

Emerged and mated females of *E. mundus* were released (one female/ plant). After four days, from release of *E. mundus* on tomato plants of 32 groups the plants were sprayed by the treatments . Then, *B. Tabaci* nymphs infested (tomato plants leaves) of each treatment were counted by stereomicroscope, then put in a glass tube and observed daily for the emergence of parasitoid adults. The number of parasitoid adults and date were scored , counted and parasitism % was calculated .

Data were subjected to analysis of variance and differences among means were significantly compared according to Duncan s Multiple Range Test (Duncan, 1955).

RESULTS AND DISCUSSION

Curative effect of crude extracts of nine aromatic plants, silica nanoparticles and insecticide (Admire) on *Bemisia tabaci*:

Unhatching egg percentage :

The obtained results arranged in Table (2) during 2016 season, percentage of unhatched *B. tabaci* eggs was influenced by different concentrations of different treatments compared to untreated tomato plants (Table 2). The highest *B. tabaci* unhatched eggs was 84.40 % on tomato plants treated by the highest concentration of geranium extract, followed by 75.20 % in tomato plants sprayed by the highest concentrations of peppermint plant extract compared with tomato plants untreated and other treatments. However, the lowest unhatched *B. tabaci* eggs was 8.00 % in tomato plants treated with low concentration of sweet basil extract compared with the other treatments (Table 2).

percentage of unhatched *B. tabaci* eggs during 2017 season was similar to that of 2016 season (Table 2).

Table 2.	The average percentage of unhatched eggs of
	Bemisia tabaci exposed to aromatic plant
	extracts, silica nanoparticles and Imidacloprid
	on tomato plants under semi natural conditions
	during 2016and 2017 seasons

	Rate ml /	Average percer	ntage of unhatched			
Treatment	100 l.	eggs				
	water	Season 2016	Season 2017			
	50	41.00 e	41.25 e			
Geranium	100	60.00 c	60.50 c			
	150	84.40 a	84.60 a			
	50	28.00f	28.25 f			
Peppermint	100	49.80 d	50.00 d			
	150	75.20 b	75.25 b			
	50	15.40 h	15.66 h			
Spearmint	100	23.80 g	24.00 g			
1	150	42.20 e	42.25 e			
	50	8.00i	8.20 i			
Sweet basil	100	10.40 i	10.50 i			
	150	15.20h	15.25 h			
	50	10.00 i	10.25 i			
Hot pepper	100	15.40h	15.50 h			
1 11	150	28.80f	29.00 f			
	50	19.20g	19.25 g			
Chris's	100	42.00e	42.20 e			
	150	64.40c	64.50 c			
	50	19.40g	19.55 g			
Rosemary	100	24.80 g	24.90 g			
5	150	29.20 f	29.25 f			
	50	22.20 g	22.25 g			
Spurge	100	51.40 d	51.44 d			
1 8	150	70.80 c	70.90 c			
	50	10.40 i	10.55 i			
Moringa	100	14.20 h	14.30 h			
0	150	23.80 g	24.00 g			
a	100 ppm	9.40I i	9.60 i			
Silica	200 ppm	21.20 g	21.32 g			
nanoparticles	300 ppm	39.80 e	39.88 e			
Imidacloprid	25	19.40g	19.55g			
Control		5.20k	5.25k			

Means followed by the same letters are not significantly different at the 5% level by DMRT

Early stages mortality percentage:

Data represented in Table (3) showed that, during 2016 season, the curative effect of the nine extracts of aromatic plant species on B. tabaci early nymphs infesting tomato plants is presented in Table (3). Mortality of *B. tabaci* early nymphs was influenced by different treatments and different concentrations compared with control (untreated tomato plant). The highest mortality of *B tabaci* early nymphs was 91.55 % in tomato plants treated with the insecticide, followed by 90.99 and 88.17 % on tomato plants sprayed with the highest concentration of silica nanoparticles and Spurge extract, respectively. However, the lowest mortality of B. tabaci early nymphs was 18.21 % on tomato plants treated with the lowest concentration of sweet basil extract followed by 24.76 % in tomato plants sprayed with the lowest concentration of hot- pepper extract (Table 3). Mortality of *B. tabaci* early nymphs during 2017 season was similar to that of 2016 season (Table 3).

The late pupal stage mortality percentage:

Data in Table (4) showed that the effect of nine aromatic plant extracts, Silica nanoparticles and admire insecticide on *B. tabaci* adults stage was influenced by different treatments and different concentration of treatments Table 3. The mortality average percentage of early stages of *Bemisia tabaci* after exposure to medicinal plant extracts, silica nanoparticles and Imidacloprid under semi natural conditions during 2016 and 2017 seasons

au	ring 2016 and	1 201 / seasons				
_	Rate ml/100	Mortality average percentage of				
Treatment	l.water –	nymphal stage				
		Season 2016	Season 2017			
	50	61.20 f	61.75 f			
Geranium	100	70.40 e	70.60 e			
	150	85.80 b	90.00 b			
	50	59.00 f	59.25 f			
Peppermint	100	67.20 e	67.20 e			
**	150	79.80 c	80.00 c			
	50	50.00 h	50.00 h			
Spearmint	100	61.20 f	61.50 f			
	15	68.40 e	68.75 e			
	50	18.21 k	18.50 k			
Sweet basil	100	41.00 i	41.25 i			
	150	55.40 g	55.75 g			
	50	24.76 j	25.00 j			
Hot pepper	100	51.40ĥ	51.50 ĥ			
	150	69.40e	69.70 e			
	50	54.80g	55.00 g			
Christ's	100	62.40 f	62.75 f			
	150	75.60d	75.75 d			
	50	51.40h	51.75 h			
Rosemary	100	68.60e	68.66 e			
•	150	79.20c	79.25 c			
	50	61.60 f	61.70			
Spurge	100	80.80 c	81.20 c			
	150	88.17a	88.25 a			
	50	51.20 h	51.50 h			
Mooring	100	60.80 f	61.00 f			
-	150	70.40 e	70.55 e			
Silica	100ppm	76.20 d	76.60 d			
	200ppm	88.40 a	88.70 a			
nanoparticles	300ppm	90.99 a	91.00 a			
Imidacloprid	25	91.55 a	91.50 a			
Control	-	2.201	2.191			

Means followed by the same letters are not significantly different at the 5% level by DMRT

Table4. The percentage of Bemisia tabaci adult
emergence failure after exposure of pupae to
number of plant extracts, silica nanoparticles
and insecticide under semi natural conditions
during 2016 and 2017seasons

Rate % of <i>Bemisia tabaci</i> adults emer Treatment ml / 100 failure				
Treatment	ml / 100 l.water	Season 2016	Season 2017	
	50	68.77 d	69.20 d	
Geranium	100	79.75 b	79.77 b	
ooraniani	150	86.39 a	87.00 a	
	50	58.67 e	58.75 e	
Peppermint	100	69.75 d	70.00 d	
	150	79.78 b	80.20 b	
	50	49.75 g	50.20 g	
Spearmint	100	56.77 f	57.00 f	
	150	62.25 e	62.75 e	
	50	13.671	14.001	
Sweet basil	100	25.75 j	26.25 j	
	150	38.75 i	39.40 i	
	50	19.25 k	19.70 k	
Hot pepper	100	33.75 i	34.25 i	
	150	44.75 h	45.50 h	
	50	48.75 g	49.00 g	
Christ's	100	69.50 đ	70.25 d	
	150	84.67 a	85.70 a	
	50	25.00 j	25.25 j	
Rosemary	100	47.75 g	48.75 g	
	150	69.75 d	70.60 đ	
	50	73.75 с	74.25 c	
Spurge	100	81.50 b	81.75 b	
	150	87.01 a	87.66 a	
	50	34.25 I	34.501	
Mooring	100	43.75 h	44.25 h	
-	150	62.5 e	62.75 e	
Silica	100ppm	69.50 d	70.00 d	
nanoparticales	200ppm	81.75 b	82.25 b	
nanoparticales	300ppm	87.25 a	87.75 a	
Imidacloprid	25	88.50 a	88.25 a	
C0ntrol	-	0.75 m	0.77 m	

Means followed by the same letters are not significantly different at the 5% level by DMRT During 2016 season, the pupal mortality was influenced by different treatments and different concentrations compared to tomato plants untreated. The highest adult emergence failures were 88.50, 87.10, 86.39 and 84.67 % in tomato plants treated with the insecticide, the highest concentration of silica nanoparticles, geranium and spurge extract, respectively (Table 4). However, the lowest of % adult emergence failure was 13.67 % in tomato plants treated with the highest concentration of sweet basil extract (Table 4). Moreover, the pupal mortality of *B. tabaci* during 2017 season was similar to that of 2016 season (Table 4).

Adult mortality percentage :

Results in Table (5) showed that the highest adult mortality of *B. Tabaci* was 80.66 % in tomato plants treated with the highest concentration of silica nanoparticle . However, the lowest *B. tabaci* adult mortality was 5.00 % in tomato planted treated with the lowest concentration of sweet basil crude extract during 2016 season (Table 5) . The pupal mortality of *B. tabaci* during 2017 season was similar to that of 2016 season (Table 5).

Table 5. Mortality percentage of Bemisia tabaciadults stage exposure to plant extracts, silicananoparticle and imidacloprid on tomatounder semi natural conditions during 2016and 2017 seasons

an	a 2017 seaso	ons			
Treatment	Rate ml/ 100 Adult mortality percenta				
I reatment	l.water	Season 2016	Season 2017		
	50	30.33 i	30.50 i		
Geranium	100	60.00 d	60.25 d		
	150	75.33 b	75.60 b		
	50	21.67 j	22.00 j		
Peppermint	100	50.67 f	50.90 f		
••	150	75.00 b	75.20 b		
	50	23.33 j	23.60 j		
Spearmint	100	33.33 i	33.50 i		
-	150	55.33 e	55.40 e		
	50	5.001	5.251		
Sweet basil	100	10.33 k	10.50 k		
	150	18.33 j	18.60 j		
	50	19.33 j	19.51 j		
Hot pepper	100	33.33 i	33.41 i		
	150	45.00 g	45.20 g		
	50	30.00 i	30.25 i		
Christ's	100	51.33 e	51.42 e		
	150	76.67 b	79.91 b		
	50	18.67 j	18.85 j		
Rosemary	100	30.00 i	30.11 i		
	150	69.33 c	69.55 c		
	50	40.67 h	40.77 h		
Spurge	100	60.75 d	60.85 d		
	150	76.25 b	76.33 b		
	50	20.00 j	20.22 j		
Mooring	100	39.67 h	39.91 h		
	150	45.33 g	45.49 g		
Silica	100 ppm	45.00 g	45.22 g		
nanoparticles	200 ppm	73.33 b	73.66 b		
nanoparticles	300 ppm	80.66 a	80.90 a		
Imidacloprid	- 25 ml	77.25 a	77.35 a		
Control	25 ml	1.251	1.331		

Means followed by the same letters are not significantly different at the 5% level by DMRT

Repellency tests:

Results presented in Table (6) recorded that, the extracts of geranium, peppermint, christ's and rosemary showed repellency effects against adults of *B. tabaci* compared with untreated plants. The leaves treated with silica nanoparticles were more attractive to *B. tabaci* than untreated leaves during 2016 season. On the other hand, the repellency effect of plant extracts, silica nanoparticle and Imidacloprid against *B. tabaci* during the season 2017 was similar to that of 2016 season.

Tuestanout	Rate ml /	Whi	Whiteflies 3 hr. post application				iteflies 24 h	r. post applicat	tion
Freatment	100 l.	Season	2016	season	2017	Seaso	n 2016	Seaso	n 2017
	water	Un treated	Treated	Un treated	Treated	Un treated	Treated	Un treated	Treated
	50	50.00	0.00a	50.00	0.00a	50.00	0.00 a	50.00	0.00 a
Geranium	100	50.00	0.00a	50.00	0.00a	50.00	0.00 a	50.00	0.00 a
Geranium	150	50.00	0.00a	50.00	0.00a	50.00	0.00 a	50.00	0.00 a
	50	49.00	1.00a	48.80	1.20a	48.20	1.80 a	48.20	1.80 a
Peppermint	100	50.00	0.00a	50.00	0.00a	50.00	0.00 a	50.00	0.00 a
	150	50.00	0.00a	50.00	0.00a	50.00	0.00 a	50.00	0.00 a
	50	44.80	5.20b	44.60	5.40b	43.60	6.40 b	43.60	6.40 b
Spearmint	100	47.40	2.60a	47.20	2.80a	48.80	1.20 a	48.80	1.20 a
*	150	49.40	0.60a	49.00	1.00a	49.40	0.60 a	49.40	0.60 a
	50	26.60	23.40f	26.20	23.80f	24.80	25.20 d	24.80	25.20d
weet basil	100	35.80	14.20e	35.40	35.40e	34.60	15.40 c	34.60	15.40 c
	150	40.00	10.00d	39.60	10.40d	40.00	10.00 b	40.00	10.00b
	50	35.40	14.60e	35.00	15.00e	34.20	15.80 c	34.20	15.80 c
lot pepper	100	39.60	10.40d	39.20	10.80d	37.80	12.20 c	37.80	12.20 c
1 11	150	41.40	8.60c	41.20	8.80c	41.8	8.20 b	41.8	8.20 b
	50	47.20	2.80a	47.00	3.00a	48.80	1.2 a	48.80	1.2 a
Christ's	100	50.00	0.00a	50.00	0.00a	50.00	0.00 a	50.00	0.00 a
	150	50.00	0.00a	50.00	0.00a	50.00	o.00 a	50.00	o.00 a
	50	42.20	7.80c	42.00	8.00c	41.80	8.20 b	41.80	8.20 b
Rosemary	100	46.20	3.80a	46.00	4.00a	46.00	4.00 a	46.00	4.00 a
2	150	49.80	0.20a	49.00	1.00a	48.20	1.80 a	48.20	1.80 a
	50	46.20	3.80b	46.50	3.50b	47.20	2.80 a	47.20	2.80 a
purge	100	48.60	1.40a	49.00	1.00a	49.20	0.80 a	49.20	0.80 a
1 0	150	49.8	0.20a	50.00	0.00a	50.00	0.00 a	50.00	0.00 a
	50	39.40	10.60d	40.00	10.00d	37.80	12.20 c	37.80	12.20 c
Aooring	100	42.40	7.60c	42.80	7.20c	41.20	8.80 b	41.20	8.80 b
0	150	47.00	3.800ba	47.50	2.50a	48.40	1.60 a	48.40	1.60 a
	100	7.00	43.00h	7.60	4.40 h	15.00	35.00 f	15.00	35.00 f
lilica nanoparticles	200	21.00	29.00g	21.80	29.25g	25.60	24.40 d	25.60	24.40d
1	300	30.60	19.40f	31.00	19.33f	37.00	13.00 c	37.00	13.00 c
midacloprid	25 ml	12.80	37.20h	12.50	37.50h	13.20	36.80h	13.20	36.80h

Fable 6. Repellency effect of plant extracts, silica nanoparticle and Imidacloprid against <i>Bemisia tabaci</i> adults dur	ing
2016 and 2017 seasons	U

Means followed by the same letters are not significantly different at the 5% level by DMRT

Effect of crude extracts of nine aromatic plant species, silica nanoparticle and Imidacloprid on the parasitism percentages of *Eretmocerus mundus* under semi-natural condition

Results illustrated in Table (7) revealed that the highest parasitism (93.88 %) was recorded in tomato plants treated with the highest concentration of hot pepper extract, followed by 86.23 % on that sprayed by the highest concentration of sweet basil extract. However, the lowest parasitism 16.53 % was obtained in tomato plants treated with admire. The date indicated generally that spraying crude extracts of these aromatic Table 7. ffect of crude extracts of aromatic plants plant species augmented the parasitism percentage of E. *mundus* than tomato plants treated with the insecticide during 2016 season. These results reveled that hotpepper extract induced the highest parasitism. However, tomato plant sprayed with spurge extract had the lowest parasitism during 2016 season (Table 7 & 8).

Effect of crude extracts of aromatic plants silica nanoparticles on the parasitism percentages of *Eretmocerus. mundus* under semi natural condition during 2017 season was similar to that of 2016 season (Table 7 & 8).

ıble	7.	ffect	of	crude	extracts	of	aromatic	plants,	silica	nanoparticles	and	imidacloprid	on	the	parasitism
	ı	hercei	ntac	zes of F	retmoceri	15	<i>mundus</i> un	der sem	ni natu	ral condition di	irino	2016 season			•

reatments	Rate ml / 100	Total no. of <i>B.tabaci</i>	Total no. of	Parasitism	Emergency
catinents	l water	nymphs	E. mundus	93.88 a	(day)
at pappar	150	245	230	93.88 a	4 - 8
ot-pepper stract	100	250	200	80.00 b	4 - 10
liaci	50	268	203	75.75 b	6 - 13
weet basil	150	247	215	87.04 a	4 - 9
tract	100	271	213	79.17 b	7-11
liact	50	253	190	75.10 b	8 - 12
	150	265	210	79.25 b	5 - 12
eranium	100	250	200	76.92 b	5-14
tract	50	255	181	70.98 d	7-15
	150	249	185	74.30 c	9-13
ppermint	100	250	175	69.79 d	10-14
tract	50	256	160	62.50 e	12 - 16
• .	150	253	199	78.66 b	8 - 12
earmint	100	248	182	73.38 c	9 - 15
tract	50	244	155	63.52 e	11 – 16
·	150	242	181	74.79 c	9 - 13
rist's	100	248	175	70.56 d	10 - 15
ract	50	244	150	61.48 e	10 - 17
	150	245	196	80.00 b	6 – 13
ooring	100	240	178	74.17 c	7 – 15
tract	50	243	165	67.90 d	9 – 18
	150	241	164	68.05 d	6 - 11
semary	100	245	152	62.04 e	8 – 15
tract	50	245	141	57.55 f	10 - 17
	150	242	122	50.41 g	13 - 18
urge	100	246	136	55.28 h	15 - 20
tract	50	242	152	62.81 e	17 - 22
	300ppm	249	105	42.17 i	13 - 23
ica	200ppm	243	125	51.44 g	13 - 23 11 - 21
anoparticles	100ppm	243	145	59.67 e	10 - 20
idacloprid	25	243	40	16.53 j	10 - 20 12 - 19
ontrol	25	251	130	51.79 g	8 - 17

Means followed by the same letters are not significantly different at the 5% level by DMRT

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Freatments	Rate ml / 100 l.	Total no. of <i>B.tabaci</i>	Total no. of <i>E</i> .	Parasitism	Emergency
reatments	water	nymphs	mundus	%	day
lot-pepper	150	251	235	93.63 a	4 - 8
Iot-pepper	100	255	205	80.39 b	4 - 10
	50	273	209	76.56 b	6 - 13
Sweet basil	150	250	216	86.04 a	4 - 9
Sweet Dash	100	253	200	79.05 b	7-11
	50	259	195	75.29 b	8 - 12
Geranium	150	268	214	79.85 b	5 - 12
Jeramum	100	266	204	76.79 b	5-14
	50	257	182	70.82 d	7-15
Jonnormint	150	250	186	74.40 c	9-13
Peppermint	100	249	174	69.88 d	10 - 14
	50	256	160	62.50 e	12 - 16
spearmint	150	253	199	78.66 b	8 - 12
spearmin	100	249	183	73.49 c	9 - 15
	50	244	155	63.52 e	11 - 16
Christ's	150	243	182	74.90 c	9 – 13
.nii ist s	100	250	176	70.40 d	10 - 15
	50	248	152	61.29 e	10 - 17
looring	150	246	197	80.08 b	6 – 13
Aloorning	100	241	179	74.27 c	7 - 15
	50	245	166	67.76 d	9 - 18
locomore	150	241	164	68.05 d	6 – 11
Rosemary	100	247	154	62.04 e	8 - 15
	50	245	141	57.55 f	10 - 17
	150	244	124	50.41 g	13 - 18
purge	100	247	137	55.28 h	15 - 20
	50	244	154	62.81 e	17 - 22
	300ppm	250	106	42.17 i	13 - 23
ilica nanoparticles	200ppm	244	126	51.44 g	11 - 21
	100ppm	247	146	59.67 e	10 - 20
midacloprid	25	245	42	16.53 j	12 - 19
Control		254	132	51.79 g	8 - 17

Table 8. Effect of crude extracts of aromatic plants, silica nanoparticles and imidacloprid on the parasitism	
percentages of <i>Eretmocerus, mundus</i> under semi natural condition during 2017 season.	

Means followed by the same letters are not significantly different at the 5% level by DMRT

Whitefly management has traditionally depended on the use of synthetic insecticides. However, the increasing resistance of Bemisia species to insecticides provides an impetus to integrated pest control measures , including bio pesticides and biological control combat this pest. Biopesticides are based on natural products and synthetic analogs of naturally occurring biochemical and are more acceptable than congenital pesticides because of their reputation for being less hazardous to humans and other non – target organisms (McClosky et al. 2000) . Among the bio pesticides are chemicals derived from a variety of plant families. The biological activity of plant extracts, bacteria, fungi, viruses and insects has been reported (Bazsik 1996; Macedo et al. 1997; Unicini Manganelli et al. 2005). In the present work, the crude extracts had toxicity and repellent effects of mooring, geranium, peppermint and spearmint extracts on Bemisia tabaci stages . However, the toxicity effects of hot- pepper, sweet basil, rosemary, spurge extracts, nano silica and the insecticide were tested against *B. tabaci* stages. The extracts of nine aromatic plants can be used as natural control agents . Most of these plants are widely distributed and easy grown . Furthermore, the extraction method is simple and cost-effective and the application techniques could be relatively easily designed for nonform use. Since B. tabaci transmits tomato leaf curl virus, developing new methods of control is obviously important. Many essential plant oils show a broad spectrum of activity against pest insects and growth regulatory and antivector activities (Opender et al. 2008). The essential oils of peppermint (Mentha piperita L.) and rosemary (Rosmarinus officinalis L.) have been reported for its insecticidal activities against greenhouse whitefly (Trialeurodes vaporariorum) (Aroiee et al., 2005). Rosemary oil is significantly repellent to the two- spotted spider mite (Miremallis and Isman 2006).

Therefore, recent efforts have expanded research and utilization of habitat management techniques for conservative biological control of arthropods in various systems (Landis et al.2000). Parasitoids are known to be attracted to plant extracts or volatiles (Vinson, 1975).

REFERENCES

- Al-Musa, A.; I.K. Nazer and N.S.Sharaf (1987). Effect of certain combined agricultural treatments on whitefly population. Dirasal J. (Agric.Sci.), 14 (11):127-134.
- Aroiee, H.; S. Mosapoor and H. Karimzadeh (2005).
 Control of greenhouse whitefly (*Trialeurodes vaporariorum*) by thyme and peppermint KMITL Science Journal,5 (2): 511-514.
- Blackman, R. L. (1971) . Variation in the photoperiodic response within natural populations of *Myzus persicae* (Sulzer). Bull. of Entomol.Res., 60(4): 533-546.
- Entomol.Res., 60(4): 533-546. Bozsik, A. (1996). Studies on aphicidal efficiency of different stinging nettle extracts. Anzeiger fuer Schaedlingskunde Pflanzenschutz Umweltschutz 69: 21-22.
- Browen, J. K.(1994). Current status of *Bemisia tabaci* as a plant pest and virus vector in agroecosystems whorledwide. FAO Plant Protection Bulletin, 42(1/2) : 3- 32.
- Protection Bulletin, 42(1/2) : 3- 32. Brown, J. K. and J. Bird (1992) . Whitefly- Transmitted geminiviruses in the Americas and the Caribbean Basin.. Plant Disease, 76(3):220-225.
- Byrne, D. N. ; Jr. T. S. Bellowes and M.P. Parrell (1990) . In :Gerling D, editor. Whiteflies in agricultural system .Whiteflies :their bionomics , pest status and management . Intercept Ltd.,Andover, Hants,227-261.

Duncan, D.B. (1955). Multiple range and multiple F test. Biometrics, 11: 1-42.

- Daoubi, M. ; A. Deligeorgopoulou ; A. J. Macias Sanchez ; R. Hermamdez-Galan ; P. B. Hitchcock ; J. R. Hanson and I. G. Collado (2005).Antifungal activity and biotransformation of diisophorone by *Botrytis cinerea* J. of Agric. and Food Chemistry, 53: (15) 6035-6039.
- (FAO) Food and Agriculture Organization (2009). FAOSTAT. Available: http:/faos. Fao.org [acces 31 December 2009]. 36 (2): 238-239.
- Greathed, A.H. (1986) . Host plants. p. 17-26. In M.J. W. Cock (ed.) *Bemisia tabaci* .A Literature survey whitefly with an annotated bioliography, CAB, Ascot, UK. On cotton .
- Giovannucci, E. (1999). Tomatoes, Tomato-Based Products, Lycopene, and Cancer: Review of the epidemiologic literature. Journal of the National Cancer Institute, 91 (4) : 317-331.
- Jones, W.A. and S.M. Greenberg (1998). Suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) instars for the parasitoid *Eretmocerus mundus* (Hymenoptera: Aphelinidae). Env. Entomol., 27(6): 1569-1573.
- Jones, D.R., (2003). Plant viruses transmitted by whiteflies. Eur. J. Plant Pathol., 109 (3) : 195 -219.
- Kim, H. G.; J. H. Jeon; M. K. Kim and H.S. Lee (2005). Pharmacological ectsofasaron aldehyde isolated from Acorusgram in eusrhizome. Food Science Biotechnology, 14(5): 685-688.
 Landis, D.A.; S. D. Wartten and G.M. Gurr (2000).
- Landis, D.A.; S. D. Wartten and G.M. Gurr (2000). Habitat management to conserve natural enemies of arthropod pests in agriculture . Ann. Rev.Entomol., 45:175-201.
- Mauchline, A. L.; Osborne, J. L.; Martin, A. P.; Poppy, G. M. and Powell, W. (2005). The effects of nonhost plant essential oil volatiles on the behaviour of the pollen beetle *Meligethes aeneus*. Entomol. Exp. Appl., 114(3): 181-188.
- Exp. Appl., 114(3): 181-188.
 Macedo, M.E.;R. A. Consoli; T. S. Grandi; A. M. Dos Anjos; A. B. Oliveira; N. M. Mendes; R. O. Queiroz and C. L. Zani (1997). Screening of Asteraceae (Compositae) plant extracts for larvicidal activity against *Aedes fluviatilis* (Diptera: Culicidae). Memoires Instituto Oswaldo Cruz, 92 (4): 565-570.

- McCloskey C . J. ; T. Arnason ; N. Donskov ; R. Chenier ; J. Kaminski ; G. B. Philo ; M. Markouk ; K. Bekkouche ; M. Larhsini ; M. Bousaid ; H. B. Lazrek and M. Jana (2000).Evaluation of some Moroccan medicinal plant extracts for larvicidal activity. Journal of Ethanopharmacology, 73(1-2): 293-297.
- Ethanopharmacology, 73(1-2): 293-297. Miremaillis, S. and M.B. Isman (2006) . Efficacy and persistence of rosemary oil as an acaricide against two-spotted spider mite (Acari. Tetranychidae) on greenhouse tomato . J. Econ. Entomol., 99 (6) : 2015- 2023.
- Mutanen, M.; A. M. Pajari; J. Levy; S. Walfisch; A. Atzmon; K. Hirsch; M. Khanin; K. Linnewiel ; Y. Morag; H. Salman; A. Veprik; M. Danilenko and Y. Sharoni (2011). The role of tomato lycopene in cancer prevention. In Vegetables, Whole Grains, and Their Derivatives in Cancer Prevention, Vol. 2, pp. 47-66. Springer, Netherlands
- Oliveira, M.R.V. ; T.J. Henneberry and M. Anderson (2001). History, current status, and collaborative research projects for *Bemisia tabaci*. Crop Prot., 20: 709-723.
- Opender, K. ; W. Suresh and G.S. Dhaliwal (2008). Essential oils as green pesticides : Potential and constraints. Biopestic . In., 4 (1): 63-84.
 Schuster, D.J. ; P.A. Stansly and J.E. Polston (1996).
- Schuster, D.J.; P.A. Stansly and J.E. Polston (1996). Expression of plant damage of Bemisia. P. 153-165. In D. Gerling & R.T. Mayer (eds.). Bemisia 1995. Taxonomy, Biology, Damage, Control and Management. Intercept, Andover, UK.
- Tare, V.; S. eshpande and R. N. Sharma (2004). Susceptibility of two different strains of Aedesaegypti (Diptera:Culicidae) to plant oils. J.of Econ. Entomol., 97: 1734-1736.
- J.of Econ. Entomol., 97: 1734-1736. Unicini Manganelli, R. E.; L. Zaccaro and P. E. Tomei (2005) . Antiviral activity in vitro of Urtica dioica L., Parietaria diffusa M. and K., and Sambucus nigra L. Journal of Ethnopharmacology 98: 323-327.
- Vinson, S.B. (1975). Biochemical coevolution between parasitoids and their hosts. Pp. 14-48 InP.(ed.) Evolutionary Strategies of parasitic insects and mites Plenum, New York.
- Walaa El-sayed (2013). Field evaluation of plant extracts and certain insecticides against *Bemesia tabaci*(Gennadius) on tomato plants and *Myzus persicae* (Sulzer) on pepper plants. J. of App. Sci. Res., 9(3): 2372-2377

تأثير مستخلصات بعض النباتات العطرية, والسليكآ النانو مترية والمبيد الحشرى (اميداكلوبرايد) على ذبابة القطن البيضاء والطفيل Eretmocerus mundus المرتبط بها إبراهيم فتحي خفاجي ' ، أميرة شوقي محمد ابراهيم ' و أسماء محمد علي الغباري ' ' معهد بحوث وقاية النبات - محطة البحوث الزراعية بسخا – مركز البحوث الزراعية

أ قسم الحشرات الاقتصادية – كلية الزراعة – جامعة كفر الشيخ

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